



# Analysis of ethylene oxide and 2-chloroethanol in sesame seeds and other food commodities

### January 19, 2022, 2:00pm-3:30pm EST

Ethylene Oxide (ETOX) is a gas that can be used as a fumigant on certain products intended for human consumption in order to reduce bacterial contamination, particularly salmonella. Several challenges are encountered when it comes to the analysis of ETOX and 2-CE in various food products: mainly due to the accumulation of high amounts of nonvolatile material in the liner, column, and possible interference with Acetaldehyde.

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# Mild processing of seafood—A review

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Revised: 19 October 2021

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#### Abstract

Recent years have shown a tremendous increase in consumer demands for healthy, natural, high-quality convenience foods, especially within the fish and seafood sector. Traditional processing technologies such as drying or extensive heating can cause deterioration of nutrients and sensory quality uncompilable with these demands. This has led to development of many novel processing technologies, which include several mild technologies. The present review highlights the potential of mild thermal, and nonthermal physical, and chemical technologies, either used alone or in combination, to obtain safe seafood products with good shelf life and preference among consumers. Moreover, applications and limitations are discussed to provide a clear view of the potential for future development and applications. Some of the reviewed technologies, or combinations thereof, have shown great potential for non-seafood products, yet data are missing for fish and seafood in general. The present paper visualizes these knowledge gaps and the potential for new technology developments in the seafood sector. Among identified gaps, the combination of mild heating (e.g., sous vide or microwave) with more novel technologies such as pulsed electric field, pulsed light, soluble gas stabilization, cold plasma, or Ohmic heat must be highlighted. However, before industrial applications are available, more research is needed.

#### **KEYWORDS**

Hurdle technology, Mild processing technologies, Seafood, Seafood quality, Seafood safety

#### **1** | INTRODUCTION

Fish and seafood are recognized for their health benefits and are widely accepted to be an essential part of a balanced, healthy diet (Carlucci et al., 2015; Mandal et al., 2020; Ahern et al., 2021). Despite this, surveys show that the average seafood consumption in Europe is considerably less than the recommended amount (Altintzoglou, Einarsdottir, et al., 2010; EUMOFA, 2019). Lack of knowledge, difficulty of preparation, cost, and inconvenience are often described barriers for the consumption of seafood (Altintzoglou, Hansen, et al., 2010; Govzman et al., 2021). A review by Carlucci et al. (2015) summarizes consumer purchase behavior toward fish and seafood products. This review concluded that consumers seem to appreciate new, convenient, processed fish products "when" the original characteristics are not significantly altered. However, an increasing level of processing (without further specification) caused consumers to perceive the modifications of the original product characteristics as a proportional loss of quality, safety, naturalness, healthiness, and nutritional value. This has driven the research and development of

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ready-to-eat and other convenient partially processed seafood products that comply with consumers' preferences for healthy, natural, high-quality, fresh or fresh-like products (Carlucci et al., 2015; Casini et al., 2015). In later years, several papers have reported the consumers' purchase decision and the willingness to pay for different labels such as sustainability (Zander & Feucht, 2018; Lawley et al., 2019; Maesano et al., 2019), social responsibility certifications (Del Giudice et al., 2018), and health and environmental benefits (Menozzi et al., 2020). In general, there is a consensus among today's consumers that eco-friendly and healthy labels are attractive.

Traditional food processing methods such as drying or extensive heating and salting cause nutrients and/or sensory quality deterioration. Alternatively, mild processing methods have been tested and improved and gained more interest from the industry and research (Boziaris, 2014; Özoğul, 2019; Ekonomou & Boziaris, 2021). Most of these technologies vary in multiple aspects; for instance, some have been used for centuries, whereas others are newly developed. Furthermore, the working mechanisms and the effect on food safety, shelf life, and sensory quality differ from technology to technology. A schematic presentation of mild processing technologies is presented in Figure 1. In the present review, thermal and physical nonthermal inactivation methods as well as mild inhibition methods applied to seafood products are thoroughly discussed. Due to the mild application of these technologies, applied alone, they are seldomly sufficient ensuring good food safety and shelf life. Hence, a combination of two or more technologies are often applied either simultaneously or consecutively, an approach known as hurdle technology (discussed in Section 4).

Knowledge regarding the potential impacts, applications, and limitations of these technologies has not previously been gathered in one place, especially with a focus on fish and seafood application. Hence, this review aims to provide an overview of existing mild postharvest processing technologies and how these can be combined (hurdle technology), including a description of working mechanisms and obtained results and evaluating each technology's applicability.

#### 2 | METHODS

Technologies included in this review were chosen based on a systematic literature search (Mandal et al., 2020) in Web of Science using the following keywords: lightly/mildly/minimally processed food, light/mild/ minimal processing technologies, nonthermal/minimal processing, or emerging/new/trends technologies/processing. In total, more than 75,000 unique publications were identified. Repeating the search by filtering seafoodrelated research, the number of unique publications was reduced to less than 6000, from which 327 met the criteria for the review process. No age restrictions were applied in the search, but for most technologies, the included studies focus on the last 5–10 years of research.

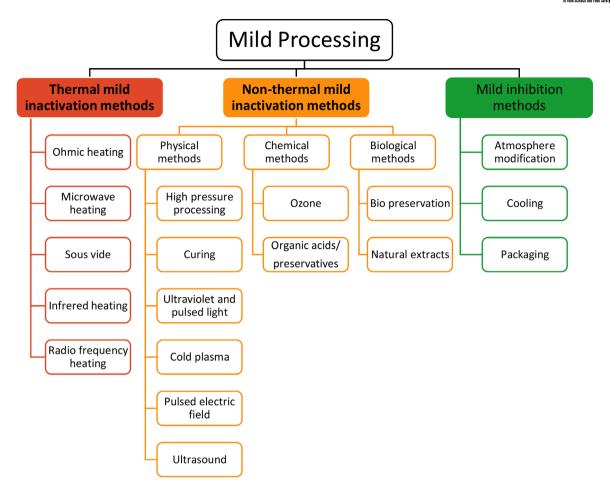
#### 3 | MILD PROCESSING METHODS FOR SEAFOOD

Many published articles use the term lightly processed, or some variant thereof (minimally processed, mild preservation, etc.), but almost all fail to define what is meant by choice of terminology. Articles that do offer an explanation or definition are seldomly in agreement. The most commonly used explanation is nonthermal processing methods (Allende et al., 2006; de Oliveira et al., 2019; Mañas & Pagán, 2005). However, this definition eliminates technologies such as sous vide, which by many is considered the exemplification of mild processing. A more nuanced definition includes maximum temperatures applied, but even those tend to vary significantly, from "[...] mild temperature; <40°C" (Barba et al., 2017) (p. 20) to "[...] temperatures remain under 100°C" (Rodgers, 2016) (p. R2309). Rajkovic et al. (2009) (p. 889) use "sublethal" to describe mild processing and extends on it by writing "[...] socalled mild decontamination [...] treatments inactivate only a part of the present microbial population [...]." This explanation only takes in part of the issue the microbiology, forgetting about all the other aspects that make up a food product. This is included in the article by Jermann et al. (2015) (p. 14) that explains it as methods that are "[...] extending product shelf life without affecting the nutritional content, organoleptic attributes and products specification." Guillou and Membré (2019), Nierop Groot et al. (2019), and Timmermans et al. (2014) use similar descriptions. In this article, the following definition of mildly processed seafood will be used:

Mild processing methods extend product shelf life and food safety by, partly or totally, inhibiting spoilage and pathogenic microorganisms and/or enzymes while affecting organoleptic attributes, nutritional content, and product characteristics as little as possible.

The mild processing concept was developed to preserve fresh quality; however, the particular focus has been to secure food safety and product shelf life due to the mild processing condition.

Broadly speaking, these mild processing methods can be separated into two different groups: thermal and nonthermal inactivation methods, although it might be argued that some processing technologies may fall under both categories. Furthermore, an additional group of mild inhibition methods may be considered as not all suitable



**FIGURE 1** Examples of different approaches to mild processing of food separated based on working mechanisms. This review article focuses on thermal and physical nonthermal inactivation methods as well as mild inhibition methods applied to seafood products

methods exhibit inactivation effects. With the growing interest in developing and refining new production, the number of potential methods is endless. Therefore, the following work will focus on mild thermal, and nonthermal physical, and chemical technologies, either used alone or in combination (hurdle technology, discussed in Section 4), to obtain safe seafood products with good shelf life and preference among consumers. Nutritional aspects, and the preservation of seafood by additives, including biopreservation will not be discussed in this paper.

#### 3.1 | Thermal mild inactivation methods

Thermal processing is the best-known preservation method and still dominates the food processing industry. The core inactivation mechanism in most thermal processing methods is the heat denaturation of the present microbiota, whereas the main difference is how the heat is applied. Besides the most used thermal processing technologies described in detail below, this group also includes radiofrequency heating, infrared heating, and Shaka technology (an agitated thermal process promoting heat transfer by forced convection).

#### 3.1.1 | Ohmic heating

The concept of ohmic heating (OH) was first reported in the late 1800, followed by the first industrial application in the 1920s, where it was used for processing milk (Prescott, 1927). The technique has been known and researched for years, but the applications of OH are relatively new (Kumar, 2018).

Two mechanisms explain OH's effect: the thermal effect, which is the most dominant, and permeability enhancement, known as electroporation (Makroo et al., 2020). The thermal effect of OH is similar to that of conventional thermal inactivation. The difference between OH and conventional heating is that the heat is produced directly within the food itself during OH. When an electric current pass through an object, the object's electrical resistance causes electrical energy to be converted to heat, an effect known as Joule heating. This is why OH is also known as Joule

Comprehensiv



heating or electrical resistance heating (Sastry, 2008). The working mechanism of OH causes an inside-out heating pattern resulting in rapid and uniform heating. Hence, no large temperature gradients will exist in the product, preventing overheating the surface, which preserves the sensory attributes of the treated food product (Tian et al., 2018). Electroporation occurs as high-voltage pulses induce and alter different electrical potentials between each side of the cell membranes. This causes hydrophilic pores and opening of transmembrane protein channels (Gómez et al., 2019; Yogesh, 2016). These pores allow intracellular content to leak from the cell, consequently losing cellular activity. The pores formed can be either reversible or irreversible depending on the extent of the treatment (Gómez et al., 2019).

The antimicrobial effect of OH has been studied intensively in products such as milk and juices (Tian et al., 2018); however, studies on seafood are limited. Bastías et al. (2015) found no difference in the microbial load of Chilean blue mussel (*Mytilus chilensis*) after being heated to a core temperature of 50, 70, or 90°C using either OH or conventional water bath. Furthermore, OH effectively inactivate *Listeria monocytogenes* in other food products (Makroo et al., 2020).

Experiments on shrimps (*Litopenaeus vannamei*) have found that processing with OH resulted in the same texture as conventional steam cooking. However, unlike steaming, OH achieved the core temperature faster  $(40 \pm 1 \text{ s com-}$ pared to  $59 \pm 2 \text{ s}$ ), and the temperature gradient within the shrimp was much smaller, regardless of the size of the shrimp or body part (head, body, or tail) (Lascorz et al., 2016). Similar findings were reported by Pedersen et al. (2016). This highlights the main advantage of OH compared to conventional heat treatment; it is possible to obtain the same characteristics but faster and without burning the surface.

Rajasekaran et al. (2021) reported OH at high voltage and short processing time (5 min to aim a core temperature of 72°C) to be beneficial regarding physiochemical and organoleptic quality. Moreover, they reported a reduction in total viable counts (TVC) up to 69% compared to untreated Green mussels (Perna viridis). The potential of high-frequency OH (20-50 kHz) has recently been highlighted, showing good water retention, low shrinkages, and beneficial textural properties of heat-treated scallops (Llave et al., 2018), as well as improved electrical conductivity during thawing of frozen tuna (Liu et al., 2017). Faster heating rates, such as those obtained through highfrequency OH, have been found protecting denaturation of actine (Llave et al., 2018), one of the major structural proteins influencing a product's water holding and textural properties. This highlights the potential of high-frequency OH to reduce the degradation of nutrients and to improve

the retention of, for example, vitamins in seafood. However, to design a thermal process operation, the knowledge of temperature distribution within the processing unit and the thermal behavior of the product is required. Such knowledge can be obtained by developing threedimensional models to predict the heating patterns of the product such as shown by Jin et al. (2020) for yellowtail (*Seriola quinqueradiata*) fillets.

#### 3.1.2 | Microwave heating

For many years, microwave (MW) ovens have been an essential appliance in most kitchens. Typically, they have been used for reheating already processed foods, but nowadays, they are used more frequently to cook raw foods both at home and in the industry.

MW technology uses electromagnetic waves, which affect the treated material in different ways. The varying electric field produced by the waveform causes dipolar molecules, such as water to oscillate back and forth. Due to the high frequencies applied in MW heating, this oscillation will occur several million times per second. Due to internal friction, these oscillations lead to volumetric heating of the food product. Second, as described for OH, when an electric current pass through an object, the object's electrical resistance causes electrical energy to be converted to heat (Chandrasekaran et al., 2013). MWs are part of the electromagnetic spectrum with a frequency range between 300 MHz and 300 GHz. The operating frequency for domestic appliances is typically 2450 MHz, whereas, for industrial applications, it is either 915 or 2450 MHz (Rosnes et al., 2011). Nevertheless, the frequencies vary in different regions of the world (Orsat et al., 2017).

MW pasteurization has mainly been applied to liquid food, such as milk and juices (Salazar-González et al., 2012), whereas the application in seafood is limited. A literature review by Tocmo et al. (2014) reported that only two studies were available regarding the inactivation of foodborne pathogens in seafood products. Sheen et al. (2013) evaluated the effect of MW cooking of catfish (Sciades herzbergii) fillets and found that 2 min at 1000 W were sufficient to eliminate (>5 log colony forming units [CFU]/g reduction) Salmonella spp. Similar results were obtained for Escherichia coli O157:H7 and L. monocytogenes at 750 and 875 W, respectively. Huang et al. (1993) reported that MW heating to a core temperature of 60°C resulted in up to 4-log CFU/g reduction in inoculated L. monocytogenes. In contrast, Aeromonas hydrophila was reduced by 5 log CFU/g. To the best of our knowledge, only one study in the past 5 years has investigated the specific effect of MW on foodborne pathogens in fish products concluding that even though MW had a significant effect on the microbial

inactivation of *L. monocytogenes*, *Staphylococcus aureus*, and *E. coli* O157:H7, a treatment above 70°C core temperature was needed to ensure complete destruction of these pathogens (Ulusoy et al., 2019).

The fact that MW heats food products faster than conventional heating underlines the main advantage of MW heating; shorter heating and exposure times are less destructive (Thostenson & Chou, 1999). This has been demonstrated for Atlantic salmon color (Lerfall et al., 2018) and nutrient content (Ersoy & Özeren, 2009). Lerfall et al. (2018) found MW heating to be beneficial compared to conventional pasteurization for the Atlantic salmon color visualized by a darker (reduced  $L^*$ ), more reddish (increased  $a^*$ ), and yellowish (increased  $b^*$ ) color. Moreover, MWtreated samples were slightly softer, but no differences were observed regarding the consumers' acceptability. In the study by Ersoy and Özeren (2009), the effect of different cooking methods on the mineral and vitamin content of African Catfish was studied. MW cooking (2450 MHz, 4 min) gave better retention of minerals and vitamin E compared to those baked in the oven (200°C, 15 min), in the grill oven (200°C, 10 min), or fried (200°C, 4 min). However, accounting for all results, the authors concluded the grilling method to be most suitable for heat processing of catfish due to better retention of vitamins (in total).

The applications of MW are versatile and include drying, blanching, baking, and extraction; however, industrial use of MW is limited (Orsat et al., 2017). This is partially explained by a series of limitations to the use of MW. MW only offers a limited penetration depth, which according to Metaxas and Meredith (1988) is about 10–20 mm at 2450 MHz. Furthermore, MW has been associated with significant uneven heating causing the formation of cold and hot spots leading to insufficient treatment or burns, respectively (Rosnes et al., 2011). The research focusing on these problems is growing rapidly, and the development of commercial systems for food processing has started (Brody, 2012; Orsat et al., 2017; Rosnes et al., 2011).

#### 3.1.3 | Sous vide

Sous vide is defined as "Raw materials or raw materials with intermediate foods that are cooked under controlled conditions of temperature and time inside heat-stable vacuumized pouches" (Schellekens, 1996) (p. 256). Sous vide was first developed in 1974, yet the technique did not reach academic research until the 1990s, and later in the mid-2000s, it became widely known to ordinary people. From there on, it has been a fast journey, with sous vide cooking equipment becoming standard appliances in many restaurants and private kitchens (Baldwin, 2012; Gonzalea-Fandos & Laorden, 2020). Despite sous vide relying on the same heating principle as traditional cooking, it differs in two fundamental ways: The raw ingredients are vacuum packaged in heat-stable plastic pouches, and second, the food is cooked using precise temperature control, often at low temperatures for a long time (Gonzalea-Fandos & Laorden, 2020). Both steps offer a range of advantages over traditional cooking. Vacuum packaging eliminates the risk of cross- or recontamination after the heat treatment, prevents the evaporation of water and loss of volatile compounds during the cooking, and reduces the access of oxygen (O<sub>2</sub>), limiting lipid oxidation and reducing the growth of a series of aerobic bacteria. The precise temperature control offers increased reproducibility and potential to pasteurize food even at low temperatures (Baldwin, 2012).

Being developed in a restaurant, sous vide was first used as a cook-serve method, with no need for storage. As sous vide has shifted from a restaurant to an industrial application, there is an increased need for prolonged storage (distribution, selling, consumers' home, etc.).

The main advantage of sous vide cooking is the benefits to the sensory and nutritional quality. Sous vide cooking of fish cakes increased the sensory shelf life (based on juiciness, tenderness, flavor, appearance, and hedonic appeal) from 4 to 16 weeks compared to conventional cooking (Shakila et al., 2009). Similarly, sous vide cooking at 70°C for 10 min significantly increased the sensory shelf life of Atlantic bonito (Sarda sarda) while maintaining food safety (Mol et al., 2012). Furthermore, Díaz et al. (2011) found no detectable rancidity associated with lipid oxidation in sous vide cooked Atlantic salmon during storage. This is in agreement with other findings showing a delay in the onset of lipid oxidation due to sous vide treatment (Díaz et al., 2009; Schellekens, 1996). Additionally, Nishioka et al. (2011) showed no loss of vitamin  $B_{12}$  during sous vide cooking of herring (Etrumeus teres), whereas traditional cooking resulted in a loss of up to 62% of the content. The sensory and nutritional quality impact of sous vide cooking has been reviewed by Creed (1995).

The gentle heat treatment applied during sous vide cooking is insufficient to render food products safe for storage. Thus, the safety of sous vide cooked and chilled products relies heavily on fast cooling and maintaining cold storage (below 4°C) for the duration of the shelf life (Baldwin, 2012). Sous vide cooked and chilled mussels (*Mytilus galloprovincialis*) (85°C for 10 min) obtained a 50% increase in shelf life compared to conventional cooking (Bongiorno et al., 2018). Similar results were obtained on fish cakes (Shakila et al., 2009). On the other hand, González-Fandos et al. (2004) showed that although 90°C (15 min) was sufficient to injure the present microorganisms thermally, they were able to recover and multiply during refrigerated storage, especially under mild temperature abuse (10°C). This

highlights the importance of proper cold storage. The shelf life of sous vide cooked and chilled fish products ranges from 6 to 42 days (Sampels, 2015).

Despite cold storage, various pathogens are still of concern for sous vide cooked fish products. Nonproteolytic *Clostridium botulinum* is one of these concerns as *C. botulinum* can form spores and survive after low or inadequate heat treatment and later start growing and producing toxins during refrigerated temperatures (Gonzalea-Fandos & Laorden, 2020). Experiments have shown that heat treatment in the temperature range of 65–90°C, including the typical temperature range of mild sous vide processing, has little effect on spores of nonproteolytic *C. botulinum*. Hence, the most efficient measure in the control of *C. botulinum* is fast cooling and refrigerated storage. Although *C. botulinum* can recover and grow at refrigerated temperature, the growth rate is slow under such conditions (Garcia et al., 1987).

Another concern is the survival and growth of L. monocytogenes. Generally, a heat treatment at 70°C for more than 2 min is recommended to ensure a 6-log reduction of L. monocytogenes (Baldwin, 2012). However, a 70°C heat treatment is seldomly applied to fish products due to the risk of protein precipitation at high temperatures. Recently, there has been an increase in low-temperature sous vide cooked seafood (42-60°C) (Gonzalea-Fandos & Laorden, 2020). However, there is a lack of data regarding the thermal inactivation of vegetative pathogens at such low temperatures (Stringer et al., 2012). This is one reason for recommending (re)heating of sous vide fish products before consumption. A sous vide treatment could potentially be combined with several other technologies such as pulsed electric field (PEF), pulsed light (PL), or soluble gas stabilization (SGS). Scant information about the combined effect of sous vide and PEF/PL is available on seafood. However, the combined effect of sous vide and SGS is discussed in Section 4.

Most of the other emerging mild processing technologies have become popular because they are fast, whereas the popularity of sous vide is derived from being slow but controlled. Sous vide combines vacuum packaging, low temperature–long time heat treatment, and fast cooling and storage to achieve safe products with high nutritional and sensory quality. Sous vide is one of the mild processing techniques that already have a wide existent in the industry and homes and restaurant kitchens worldwide.

# 3.2 | Nonthermal mild inactivation methods

Nonthermal processing methods are a diverse group of technologies, most of which rely on different inactivation mechanisms. Therefore, these methods are often separated into three different subgroups: physical, chemical, and biological. Besides the most used methods described below, the physical nonthermal group also includes ionized radiation, oscillating magnetic field, PEF, and ultrasound methods. Chemical food preservation has long been a wellestablished field. However, with the increasing demand for natural food products and green labels, there has been an increase in the use of so-called biological preservations, such as competitive microbiota (bio preservation) and use of essential oils or herb extracts (Banerjee & Verma, 2015; Rosnes & Skipnes, 2017). The use of chemical and biological preservation methods is outside the scope of this review and will not be described in further detail.

#### 3.2.1 | High-pressure processing

High-pressure (HP) processing, also known as high hydrostatic pressure, uses pressure between 100 and 1000 MPa to inactivate or reduce microorganisms and enzymes to a safe level. The application of HP processing for food preservation was first tested more than a century ago when Hite (1899) reported increased shelf life of milk after pressure treatment. However, scientific development and industrial application are much newer and have evolved over the last few decades (Farr, 1990; Ekonomou et al., 2020; Ekonomou & Boziaris, 2021; Shynkaryk et al., 2020; Truong et al., 2015). Originally HP treatment was used for fruit products such as juices and jams, but the use has since expanded into almost all parts of the food processing industry. Despite several successful applications in the food industry, high equipment costs (30,000-770,000\$) limit its use in smaller and medium-sized enterprises or lowproduction-volume applications (Pinto et al., 2020; van Wyk et al., 2018). However, contracting of such equipment has gained in popularity.

Application of HP causes a decrease in product volume (Martínez-Monteagudo & Balasubramaniam, 2016). This volume change affects all cellular components simultaneously due to the isostatic principle, which states that pressure is instantaneously and uniformly transmitted throughout the sample under pressure, regardless of shape and size (Smelt, 1998). The changes in product volume are accomplished by breaking molecular interactions, especially weaker interactions such as hydrogen bond, van der Waals forces, electrostatic force, and hydrophobic interactions (Tauscher, 1995). Consequently, proteins, including enzymes, polysaccharides, and nucleic acids may be subject to alteration in structure and functionality, whereas amino acids, vitamins, flavor compounds, and other small molecules remain relatively unaffected (Patterson, 2014). The ability to cause microbial inactivation while retaining

quality showcases one of the main advantages of HP processing compared to conventional thermal treatment.

Multiple mechanisms have been suggested for the inactivation and/or death of vegetative bacterial cells caused by HP processing (Ferreira et al., 2016). These include changes in morphology, damage to the cell membrane (Ritz et al., 2002), and protein denaturation leading to changes in physiology, including synthesis of vital components and maintenance of intercellular conditions (Ferreira et al., 2016; Tholozan et al., 2000).

Regardless of the mechanisms, HP processing efficiently reduces microbial counts of various seafood species (Ekonomou & Boziaris, 2021; Truong et al., 2015). Generally, the application of 300 MPa for a few min at room temperature has been suggested as an adequate treatment to inhibit vegetative bacteria in many food products (Farkas & Hoover, 2000). This has been confirmed in multiple fish products (Erkan et al., 2010; Hurtado et al., 2000; Kamalakanth et al., 2011; Yagiz et al., 2007) as well as shellfish (Linton et al., 2003; López-Caballero, Pérez-Mateos, Bonderías, et al., 2000). Truong et al. (2015) reviewed the effect of various HP studies concerning microbial reduction and shelf life extensions. Although previous experiments agree on the overall effect, a direct comparison can be difficult. Chéret et al. (2005) and Teixeira et al. (2014) both performed experiments on sea bass (Dicentrarchus labrax), both at 400 MPa for 5 min, but obtained different bacterial reductions, 3.2 and 0.44 log CFU/g, respectively. The initial bacterial load (TVC of 6.0 and 4.4 log CFU/g in Chéret et al. [2005] and Teixeira et al. [2014], respectively) and the pressurization rate (3 and 14 MPa/s, respectively) did, however, affect the obtained results. Apart from treatment conditions, the effectiveness of HP processing on microbial inactivation is significantly affected by the characteristics of the microbiota. In general, it is assumed that Gram-negative (G<sup>-</sup>) bacteria are more sensitive to pressure than Gram-positive  $(G^+)$ . This is highlighted by the findings of Linton et al. (2003), who found G<sup>+</sup> to make up 58% of the total microbiota of mussels (Mytilus edulis) before treatment compared to 91% after HP processing (500 MPa for 2 min).

Oyster (*Crassostrea gigas*) is a seafood type for which HP processing has gained the most popularity. Both due to the ability to reduce the load of *Vibrio parahaemolyticus*, but equally important due to a series of quality aspects (Murchie et al., 2005). During HP processing, the adductor muscle of oysters detaches from the shell, opening the oyster, known as shucking. He et al. (2002) report 100% full release of adductor muscle after HP processing at 310 MPa. Similar findings were made by Rong et al. (2018). Furthermore, the HP-shucked oysters had a higher yield with fewer damages than traditionally hand-shucked oysters. Following HP processing, moisture content of the oysters had increased (Cruz-Romero et al., 2004; Rong et al., 2017), which explains the reports of more voluminous and juicy oysters after HP processing (López-Caballero, Pérez-Mateos, Montero, et al., 2000). Overall, sensory evaluation found HP-processed oysters to be more acceptable (Johnston et al., 2003) and with a lower quality index method (QIM) score (indicating fewer defects) (He et al., 2002; Yu et al., 2018).

The major drawback reported regarding the HP processing of oysters is the color change. At pressure above 300 MPa, increased whiteness ( $L^*$  value) and reduced transparency of the oysters were reported, resulting in a cooked appearance (Cruz-Romero et al., 2004). These observations also represent other molluscs (Gou et al., 2010; Hughes et al., 2016). HP-induced color changes have also been observed in HP-processed squid (Loligo bleekeri) (Nagashima et al., 1993), Atlantic cod (Gadus morhua), Atlantic salmon, mackerel (Scomber scombrus) (Christensen et al., 2017), and yellowfin tuna (Thunnus albacares) (Kamalakanth et al., 2011). Observed color changes in meat are often associated with three main mechanisms: (1) denaturation of myoglobin, (2) modification or disruption of the porphyrin ring, and (3) changes in the myoglobin redox chemistry (Bak et al., 2019). However, in seafood, increased lightness  $(L^*)$  is associated with HPinduced cold denaturation of globin and myofibril proteins. On the other side, changes in redness  $(a^*)$  are most likely affected by oxidative mechanisms (de Oliveira et al., 2017). The color of fish and shellfish products plays a vital role in consumers' perception of quality (Garber et al., 2003). These discolorations are therefore of great concern to the processing industry. Another concern regarding HP processing of seafood is the influence on water holding capacity (WHC). Decreased WHC as a function of treatment pressure (200-400 MPa) has been observed on sea bream (Sparus aurata) (Campus et al., 2010) and cold smoked Atlantic salmon (Lakshmanan et al., 2007). WHC highly depends on protein-water interaction, explaining the decrease following pressure-induced protein denaturation.

HP processing is a fast-expanding processing method gaining popularity in multiple parts of the food industry. The popularity primarily stems from the ability of HP processing to inactivate vegetative bacteria and most autolytic enzymes while causing minimal deterioration to the nutrient or sensory quality. Relying on protein denaturation as a mechanism for bacterial inactivation, it is inevitable to cause alterations to the product itself. This includes a cooked appearance due to surface protein denaturation or denaturation of color complexes and alterations to WHC, texture, and induction of lipid oxidation.



#### 3.2.2 | Pulsed electric fields

PEF is an emerging nonthermal technology with great potential for cost-effective and eco-friendly applications in the food industry (Ekonomou & Boziaris, 2021). It is widely used on liquid and semiliquid food products. However, for seafood applications, further research is needed before applications can be commercialized. The basic principle of the PEF technology is the application of short pulses (a few nanoseconds to several milliseconds) of high-voltage electric fields between two electrodes with the intensity in the order of 0.1-80 kV/cm (Barba et al., 2017). The processing time is calculated by multiplying the number of pulses times with the effective pulse duration. The research on PEF related to seafood processing has increased in popularity due to its potential to inhibit microorganisms (Shiekh & Benjakul, 2020) and altering structural properties beneficial for, for example, salting (Cropotova, Tappi, Genovese, Rocculi, Laghi, et al., 2021). However, the number of high-quality publications focusing on seafood is restricted to less than 15 studies. One of the benefits of using PEF is the low impact on the sensory characteristics. Improved sensory characteristics after a PEF treatment have been reported on freshwater mussels (Zhou et al., 2017) and Asian seabass (Chotphruethipong et al., 2019). Cropotova, Tappi, Genovese, Rocculi, Laghi, et al. (2021) reported shorter brining times and increased salt uptake when PEF was used as a pretreatment before salting. The applied intensity of the current was set at 10 and 20 A (corresponding to a field strength of 0.3 and 0.6 kV/cm) before sea bass salting in brine with 5% and 10% salt concentration, respectively. However, the combination of PEF with brine salting resulted in an increase in primary and secondary lipid oxidation products expressed as peroxide value, conjugated dienes, and 2-thiobarbituric acid reactive substances in PEF-treated samples compared to untreated ones (Cropotova, Tappi, Genovese, Rocculi, Dalla Rosa, et al., 2021). In addition to the beforementioned applications, PEF can improve the extraction of nutritional and bioactive compounds. A few studies on seafood have recently been published, for example, on Pacific white shrimp (Litopenaeus vannamei) (Gulzar & Benjakul, 2020), fishbone (He et al., 2014), and sea bass and sea bream rest raw material (Franco et al., 2020), but the number of potential applications is considerable.

#### 3.2.3 | Curing

Curing is a collective term for traditional processing methods such as drying, salting, smoking, pickling, marinating, or combinations thereof (Arason et al., 2014). Unlike most of the mild processing methods mentioned up until now, curing is not a new or emerging technology, but is in fact one of the oldest methods of preserving fish (Løvdal, 2015). According to Huss et al. (2003), cured products can be divided into four diverse groups: (1) mildly preserved seafood, including lightly salted, some marinated, and cold smoked seafood products; (2) fermented seafood; (3) semipreserved seafood, including salted and/or marinated fish and caviar; and (4) smoke-dried or heavily salted seafood products, including stockfish. The following will only focus on the physical methods within the mildly preserved seafood category in line with the overall topic.

#### Smoking

Smoked seafood includes two groups separated based on the temperature of processing: cold smoked or hot smoked. Cold smoked products are processed at temperatures below 33°C, classifying them as mildly processed (Løvdal, 2015).

A traditional cold smoking process involves salting, drying, and finally, smoking. The primary purpose of salting is to lower the water activity  $(a_w)$  to inhibit spoilage mechanisms (Sperber, 1983), and it can be done either by dry, brine, or injection salting. An additional decrease in  $a_{\rm w}$  takes place during the drying and smoking steps. The smoking step further preserves the product through the release of formaldehyde and phenols known to inhibit the growth of multiple microorganisms and limit oxidative reactions (Kjällstrand & Petersson, 2001; Varlet, Prost, et al., 2007). Analyses have identified more than 200 different substances to be released during smoking (Arvanitoyannis & Kotsanopoulos, 2013), not all of which are beneficial. Especially polyaromatic hydrocarbon (PAH) compounds such as benzo(a)pyrene are of concern due to their link with cancer development.

Cold smoked seafood is very sensitive to deterioration and based on sensory evaluations it has a limited shelf life of 3-5 weeks when stored at 4°C (Leroi et al., 2001; Løvdal, 2015; Rørvik et al., 1991). The spoilage of cold smoked products is mainly ascribed to off-flavors resulting from microbial growth and metabolism (Leroi et al., 2001; Truelstrup Hansen et al., 1996). Several studies have shown that the microbiota of cold smoked products is dominated by lactic acid bacteria in combination with other spoilage bacteria such as Photobacterium phosphoreum or Enterobacteriaceae (Leroi et al., 1998; Olofsson et al., 2007; Truelstrup Hansen & Huss, 1998). One explanation for the high variability in the microbiota of cold smoked seafood is that the spoilage is highly dependent on the processing combinations. These variations include different salting methods, salt concentrations, degree of drying, and smoking method, just to name a few. For instance, Truelstrup Hansen et al. (1996) found Brochothrix thermospacta in brine-injected samples, but not in dry-salted samples, which, on the other hand, was found to be dominated by *P. phosphoreum*. One of the biggest concerns about cold smoked seafood is the potential survival and growth of *L. monocytogenes* (Jami et al., 2014). A summary of *L. monocytogenes* prevalence in retail cold smoked Atlantic salmon is presented by Løvdal (2015), reporting results between 0% and 61%, averaging at 9.8%.

Although smoking traditionally was used to preserve and extend shelf life, nowadays it is primarily applied to develop favored sensorial characteristics. Different processing settings can have a significant impact on the quality of the final product. For instance, Martinez et al. (2012) found dry salting to result in firmer cold smoked products, compared to brine salting, in agreement with findings by Birkeland et al. (2004). The firmness is facilitated by the removal of water during the salting step. This again explains the differences in obtained yield due to different salting strategies (Birkeland et al., 2004; Birkeland & Bjerkeng, 2005; Bjørnevik et al., 2018). The additional drying of the surface obtained by dry salting also influences the surface color, which generally is found to be darker and less red (decreased  $L^*$  and  $a^*$ ) compared to brine-salted smoked Atlantic salmon (Bjørnevik et al., 2018; Lerfall et al., 2011). Various settings within each of the methods mentioned (salting time and temperature, salt concentration, etc.) play a significantly role in determining the extent of the discussed effects (Birkeland & Bjerkeng, 2005; Goulas & Kontominas, 2005).

The most important factor influencing the characteristics of smoked fish products is the smoking process itself. Smoke can be generated by a variation of pyrolytic applications (Birkeland & Skåra, 2008) or by the application of liquid smoke, also known as purified condensed smoke (PCS) (Hattula et al., 2001). The previously mentioned concern over PAHs resulting from pyrolysis is one reason for the development of PCS. Smoke condensates are usually obtained from wood smoke produced by smoldering wood chips or sawdust followed by refining and rinsing steps to remove unwanted compounds (Guillén et al., 2000; Montazeri, Oliveira, et al., 2013). Despite the filtering, the active antimicrobial compounds are still the same and research has shown an inhibiting effect of several types of PCSs (Faisal et al., 2019; Guilbaud et al., 2008; Montazeri, Himelbloom, et al., 2013). PCS is used either by dipping the seafood product in a diluted PCS solution or by atomizing it and spraying the sample in a closed chamber.

Valø et al. (2020) compared traditional cold smoking with that of atomized PCS and found atomized PCS to result in lower aerobic plate count and better growth suppression than traditional cold smoking. Moreover, the PSCprocessed salmon were firmer, darker, and slightly less reddish and yellowish than those smoked traditionally. On the other hand, de Araújo et al. (2020) found no significant differences in bacterial counts in catfish products in agree-

ment with Özpolat and Patir (2016). Varlet, Serot, et al. (2007) compared sensory evaluations of pyrolysis-smoked and PCS-produced Atlantic salmon and found the samples to be significantly different, as PCS produced salmon was classified as "grassy" and "cold" compared to "buttery" for the traditionally smoked products. In contrast, Birkeland and Skåra (2008) found no significant differences in any of the evaluated sensory traits a week after processing with either PCS or traditional cold smoking. The different findings can easily be explained by using different condensates, as they differ significantly in flavor based on different production methods (Kostyra & Barłyko-Pikielna, 2006). This also highlights the possibility to adjust the use of different PCS to obtain the wanted flavor profile (Martinez et al., 2007; Martinez et al., 2012). Regardless of PCS type used, there is some characteristic of the traditionally woodsmoked products that are hard to mimic with PCS. This includes color, where PCS results in a lighter, paler surface color (lower  $L^*$ , lower  $a^*$ , and lower  $b^*$ ) than woodsmoked fish products (Birkeland & Skåra, 2008; Cardinal et al., 2001; Hattula et al., 2001).

#### 3.2.4 | Ultraviolet and pulsed light

The use of ultraviolet (UV) light treatment to preserve foods was first discovered in the 1930s and has since become a widespread disinfection method in multiple industries. The limitations to continuous UV treatment led to the development of flash lamps as an alternative for delivering UV radiation. The use was first reported in the late 1970s; however, PL treatment for microbial inactivation first reached the scientific literature in the 1990s (Bank et al., 1990; Mandal et al., 2020). Application of both for equipment, processing plant, and packaging material disinfection has long been used, but the use for direct processing of foods is relatively new, although fast growing (Mandal et al., 2020).

UV light is light in the electromagnetic spectrum region from 100 to 400 nm. However, the UV spectrum is often divided into three types based on wavelength: UV-A with 320–400 nm, UV-B with 280–320 nm, and UV-C with 200– 280 nm, the latter having the strongest germicidal properties (Bintsis et al., 2000). The energy released during UV treatment causes the formation of DNA photoproducts, most importantly pyrimidine dimers (Lee et al., 2015). This covalent cross-link between two pyrimidines of the same DNA strand can cause interruption to both transcription and translation leading to loss of function and death of the cell (Regan et al., 1968; Sharma, 2010).

PL, also known as high-intensity, broad-spectrum pulsed light (Roberts & Hope, 2003), high-intensity pulsed UV light (Ngadi et al., 2012), intense light pulses (Gómez-López et al., 2005), intense pulsed light (Choi et al., 2010), pulsed UV light (Sharma & Demirci, 2003), or pulsed white light (Marquenie et al., 2003), uses short pulses of intense, broad-spectrum light to inactivate microorganisms. PL processing uses light between 200 and 1100 nm, thus including UV, visible, and some infrared light. The germicidal effect of PL is mainly ascribed to the effect of UV-C, as described above (Gómez-López et al., 2007).

Both methods are considered efficient for decontaminations and have been used for microbial inactivation of foods, food contact materials, air, and water (Kramer et al., 2017; Oms-Oliu et al., 2010). Cheigh et al. (2013) compared continuous UV-C and PL treatment for the inactivation of L. monocytogenes in solid medium. They found PL for up to 350 s resulted in more than a 6-log CFU/g reduction, similar to MacGregor et al. (1998) and Rowan et al. (1999). However, the penetration depth is poor for liquid media (D-value of 93  $\pm$  5 s), but a thin-profile treatment can be considered (Pollock et al., 2017). Comparably, treatment using continuous UV-C treatment for up to 1000 s resulted in a 4-log CFU/g reduction. Furthermore, the significantly longer treatment time resulted in increased sample temperature. Conversely, the same PL treatment used for L. monocytogenes-inoculated shrimps, Atlantic salmon, and flatfish (Paralichthys olivaceus) fillets resulted in approximately 2.2-, 1.9-, and 1.7-log CFU/g reduction, respectively, whereas UV-C treatment gave no significant reduction (Cheigh et al., 2013). This highlights one of the main disadvantages of using UV and PL treatment for food processing-the penetration depth. Although the penetration depth is higher for PL than continuous UV-C treatment (Oms-Oliu et al., 2010), the uneven surface of food products can harbor microorganisms that will not be affected by the treatment. Similarly, if there is a high microorganism population density, they will be shadowing each other, hindering effective disinfection (Cheigh et al., 2013). Despite this, the use of continuous UV or PL treatment has shown promising results for multiple seafood products, which includes the inactivation of E. coli O157:H7 and L. monocytogenes by UV-light, on inoculated raw salmon fillets (Ozer & Demirci, 2006), improved shelf life of vacuum packaged Colossoma macropomum × Piaractus mesopotamicus, and pirarucu (Arapaima gigas) fillets (Bottino et al., 2016; de Souza Lira Santos et al., 2018), improved microbiological stability and sensory quality of dried seafood (Lee et al., 2015), and no negative sensory changes in UV-C-treated (50 mJ/cm<sup>2</sup>) cold smoked salmon (Holck et al., 2018). However, for raw salmon fillets, doses higher than 200.0 mJ/cm<sup>2</sup> introduce unwanted organoleptic characteristics (Pedrós-Garrido et al., 2018).

The main concern regarding the use of UV-C light for food processing is that it is known to be a potent prooxidant agent (Mendes de Souza et al., 2013). Nevertheless, experiments on both lean and fatty fish species have shown that the prooxidative effect is neglectable when using mild treatment dosages (Monteiro et al., 2017; Rodrigues et al., 2016). Furthermore, only applying treatment in short pulses as in PL has shown significantly decreased oxidation rates (Heinrich et al., 2016). UV-based treatments have been shown to cause discoloration to food products, especially dark meat, fruit, and vegetables (Heinrich et al., 2016). In contrast, little or no changes in color have been reported for Nile tilapia (*Oreochromis niloticus*) (Monteiro et al., 2017), sea bass (Molina et al., 2014), Atlantic salmon, flatfish, shrimps (Cheigh et al., 2013), and dried squid (*Todarodes pacificus*) (Lee et al., 2015).

UV treatment, continuous or pulsed, has gained popularity because it does not use chemicals or leave residues, the heat is minimal, it is fast and economical, and it occupies very little space. It has been used for decontamination of air, water, and equipment for long, and applications for food products are growing. However, there is still limited research regarding seafood products, as the low penetration depth of UV radiation and absorption of energy by food constituents limit the efficiency.

#### 3.2.5 | Cold plasma

The term "plasma" refers to the fourth state of matter first discovered in 1928 (Saklani et al., 2019). It is a partially ionized gas that can be generated in two ways: (1) by heating gas to extreme temperatures (approximately 1000-10,000 K), leading to the formation of thermal plasma. The high temperature of this method renders it unsuitable for food processing (Samal, 2017). (2) Alternatively, gas is passed through a high-energy electric field, which disrupts and breaks down the gas's equilibrium state by the formation of ions and electrons. The latter is known as nonthermal or cold plasma (CP) (Kulawik & Tiwari, 2019). As the name indicates, temperatures of CP are close to ambient temperatures; hence, it does not heat the treated product (Misra et al., 2015), making the methods suitable for mild processing. There are two forms of CP, low-pressure plasma systems and atmosphere condition plasma, the latter of which is the most used due to the more accessible and cheaper utilization (Misra et al., 2011).

When gases, typically  $O_2$ , nitrogen (N<sub>2</sub>), argon (Ar), atmospheric air, or a mixture hereof, are electrified, an assortment of ions, electrons, and free radical species is generated (Olatunde & Benjakul, 2018). These molecules are responsible for microbial inactivation. The exact working mechanism of CP microbial inactivation is not fully understood, but four different suggestions have been made: (1) the production of reactive molecules, (2) UV radiation, (3) the production of charged particles, and (4) the production of ozone (Guo et al., 2015). The overall effect is probably due to a combination of two or more. A common feature for all these reactions is that they cause oxidative degradation of microbial components, including the membrane leading to microbial injury or death (Kulawik & Tiwari, 2019). This highlights the benefit of  $O_2$ in the atmosphere when working with plasma. Eto et al. (2008) and Patil et al. (2014) found that addition of  $O_2$  to the atmosphere increased the efficiency of CP decontamination. Besides gas atmosphere composition, the efficiency of CP depends on the type of plasma generation unit (dielectric barrier discharge and atmospheric pressure plasma jet are the most commonly used), the product (composition, size, and surface), CP generation parameters (voltage, frequency, and time), exposure mode (indirect or direct contact), and microorganisms present (Liao et al., 2017).

The use of CP for food processing or food packaging decontamination is relatively new, and up until recently, the application has been focusing on fresh produce (Critzer et al., 2007; Fernandez-Gutierrez et al., 2010; Perni et al., 2008). The main parts of research regarding CP processing of seafood only date back to the last couple of years (Kulawik & Tiwari, 2019). Despite promising results from fresh produce, the findings from fresh seafood samples have been discouraging. Kulawik et al. (2018) reported no significant reduction in microbial load after CP treatment of sushi at up to 80 kW, 50 kHz for 5 min. Similar reports have been made for other fish products (Albertos et al., 2017; Albertos et al., 2019; Chiper et al., 2011). Considering dried and semidried seafood products, the findings are highly different. For example, CP has been reported to cause inhibition of an array of different microorganisms, including bacteria (Choi et al., 2016, 2017; Puligundla et al., 2018), yeast, and molds (Park & Ha, 2015; Puligundla et al., 2018). A summary of the findings is presented in Kulawik and Tiwari (2019). Although reports have not been made from seafood products, CP has been shown to inactive bacterial spores in culture samples (Tseng et al., 2011).

The main disadvantage to the use of CP, and why fresh seafood has often been considered unsuitable for CP treatment, is the possibility of increased oxidation rate. As stated above, CP works by causing oxidative stress to the microorganisms; however, a similar effect has been suggested to the seafood product itself. All identified studies that investigated oxidation levels of CP-treated fresh or dried seafood reported an increase in oxidation rates (Albertos et al., 2017; Albertos et al., 2019; Choi et al., 2016, 2017; Kulawik et al., 2018; Park & Ha, 2015; Puligundla et al., 2018). The oxidation level depends on the CP treatment conditions showing increased oxidation rates due to higher voltage and holding times. A high-voltage treatment (e.g., 80 kV) will give an excellent inhibitory effect on microbial counts but also increase oxidation rates. However, lower voltage (e.g., 70 kV) and shorter treatment times (<5 min) will reduce the number of oxidation products such as peroxides and dienes (Albertos et al., 2017; Albertos et al., 2019). Similarly, replication of the same studies reported a significant decrease in moisture content following CP treatments and significant color changes, including a reduction in lightness. On the other hand, no adverse changes in sensory parameters were reported (Choi et al., 2016, 2017; Kim et al., 2015). One study even reported improved appearance, color, and overall acceptance scores following CP treatment of semidried Pacific saury (*Cololabis saira*) at 20 kV, 58 kHz for up to 10 min (Puligundla et al., 2018).

CP is a method of gaining interest from research groups worldwide because it is a cost-effective, environmentally friendly method that can eliminate microorganisms, including spores. However, applications in the food industry, especially seafood, are still scarce.

#### 3.3 | Mild inhibition methods

Most of the above technologies aim to inactivate or kill microorganisms, whereas others rely on inhibiting the microorganisms by reducing growth and propagation without eliminating the microorganisms present. The most used example of the latter is the application of carbon dioxide ( $CO_2$ ) in food processing and packaging (Figure 2).

 $CO_2$  has long been known for its bacteriostatic and antifungal effect and it has been demonstrated that  $CO_2$  can extend the growth lag phase and reduce the growth rate during the logarithmic growth phase of several bacteria (Church, 1994). Hence,  $CO_2$  is extensively used for atmosphere modification of multiple food products, including seafood.

Although the antifungal and antimicrobial effect has been demonstrated in multiple experiments (DeWitt & Oliveira, 2016; Sivertsvik et al., 2002; Stammen et al., 1990), the mechanism is not fully understood. In the beginning, it was believed that the bacteriostatic effect of CO<sub>2</sub> was solely due to the replacement of O<sub>2</sub>. However, this theory was rejected when experiments showed markedly improved bacterial inhibition when using 100% CO2 compared to 100% N<sub>2</sub> (Daniels et al., 1985). CO<sub>2</sub> is easily absorbed in most food products due to its high solubility in water and liquid lipids (Abel et al., 2018). The dissolvement of CO<sub>2</sub> facilitates a drop in surface pH due to carbonic acid formation (Knoche, 1980). Although bacteriostatic, the pH drop cannot account for the entire bacterial inhibition observed from CO<sub>2</sub> processing (Coyne, 1933). Today there is a consensus that the effect of  $CO_2$ is due to intracellular accumulation causing disruption of the normal physiological equilibrium, and four mechanisms have been identified: (1) alteration of cell membrane functions including cellular uptake and release, (2) inhibition of bacterial enzymes, (3) intracellular pH changes,



**FIGURE 2** Various forms atmosphere modification, vacuum packaging, emitter, and modified atmosphere packaging. All of these can be applied in combination with soluble gas stabilization

or (4) immediate changes in physicochemical properties of proteins (Sivertsvik et al., 2002). The bacteriostatic effect is probably a combination of all the mentioned mechanisms. The mechanisms described highlight the importance of  $CO_2$  concentration in the product, as demonstrated by Devlieghere et al. (1998a, 1998b), who found the growth inhibition of microorganisms in modified atmosphere (MA) to be determined by, and proportional to, the concentration of dissolved  $CO_2$  in the product.

#### 3.3.1 | Soluble gas stabilization

Due to the solubility of  $CO_2$  in the water and liquid lipids (Abel et al., 2018; Gill, 1988), absorption of  $CO_2$  by the product in an MA packaging system results in either a pressure reduction, volume reduction, or both, depending on the packaging material (Sivertsvik et al., 2002). This volume reduction can lead to package collapse, one of the main disadvantages of  $CO_2$  in MA packaging. In order to overcome this issue, a filler gas can be introduced, reducing the percentage of CO<sub>2</sub>, thereby lessening the volume change, but also reducing the bacteriostatic effect of the  $CO_2$ . Alternatively, a higher gas-to-product volume (g/p) ratio can be applied, typically in the range of 3:1-4:1, implicating a packages size four to five times the actual size of the product (Sivertsvik et al., 2004). The consequence is lower packaging efficiency, increased distribution costs, and an increased amount of plastic materials and waste produced. An alternative for reducing the packaging size is dissolving the  $CO_2$  into the product before packaging, a method known as soluble gas stabilization (Sivertsvik, 2000, 2003). SGS has been shown to prevent package collapse, even when low g/p ratios are applied (Birkeland & Rotabakk, 2014; Rotabakk et al., 2006; Rotabakk et al., 2008; Sivertsvik & Birkeland, 2006). SGS treatment is effectuated at low temperature and pressure equal to or above 1 atm. Because the solubility of CO2 increases at lower temperature and higher partial and/or total pressure, a sufficient amount of CO<sub>2</sub> can be dissolved into the product during 1-2 h in pure CO<sub>2</sub> (Sivertsvik et al., 2004). Despite

being designated as such, SGS is not a packaging technology by definition. Hence, SGS pretreatment is followed by repackaging after ended treatment, either in vacuum or MA packaging (Birkeland & Rotabakk, 2014). Mendes and Gonçalves (2008a) compared SGS pretreatment followed by vacuum packaging with pure vacuum-packaged sea bream and sea bass and found SGS to delay the growth of naturally present bacteria, in agreement with Mendes et al. (2011) for octopus (Octopus vulgaris). Furthermore, SGS pretreatment maintained the initial sensory characteristics and quality longer, resulting in a 2-3 days extension of shelf life than pure vacuum-packaged samples (Mendes & Gonçalves, 2008a). Most literature reporting the use of SGS does so in combination with MA packaging, as it is believed to be necessary to repack in MA after SGS treatment to maintain the effect of the dissolved CO<sub>2</sub> (Birkeland & Rotabakk, 2014). Abel, Rotabakk, Rustad, et al. (2019) found MA packaging of SGS-pretreated, pasteurized Atlantic salmon loins to significantly reduced the growth of Listeria innocua by extending the lag phase and reducing the growth rate, which was in agreement with the findings of Abel, Rotabakk, and Lerfall (2019). Reduction in bacterial growth by SGS followed by MA packaging has also been observed for shrimps (Pandalus borealis) (Sivertsvik & Birkeland, 2006), Atlantic halibut (Hippoglossus hippoglossus) (Rotabakk et al., 2008), Atlantic cod mince (Birkeland & Rotabakk, 2014), and Atlantic salmon fillets (Sivertsvik, 2003).

The relationship between measured bacterial growth and perceived quality and shelf life is not straightforward. Despite the positive effect on bacterial inhibition observed from SGS treatment, multiple experiments have shown that SGS does not provide the ability to prolong shelf life when evaluated based on sensory scores. They report no beneficial or adverse effect of the SGS treatment (Abel, Rotabakk, Rustad, et al., 2019; Birkeland & Rotabakk, 2014; Mendes & Gonçalves, 2008b; Mendes et al., 2011; Rotabakk et al., 2008). However, the studies who has reported an increase in sensory shelf life, ranging from 4 to more than 9 days, were primarily based on off-odor evaluations (Mendes & Goncalves, 2008a; Rotabakk et al., 2006; Sivertsvik & Birkeland, 2006). The effect of SGS is often ascribed to inhibition of the specific spoilage organisms and inhibition of oxidative rancidity. It has been shown that an increased percentage of CO<sub>2</sub> in the headspace can induce increased drip loss (DL) as a result of pH changes causing confrontational changes to the proteins (Davis, 1998). No such effect was observed in any of the experiments analyzing DL after using SGS (Abel, Rotabakk, Rustad, et al., 2019; Al-Nehlawi et al., 2013); some even reported a reduction in DL (Rotabakk et al., 2008; Sivertsvik & Birkeland, 2006). All the mentioned studies make a comparison between SGS-treated samples and regular MA packaging, hence all samples containing some level of  $CO_2$ . This might explain the lack of difference.

#### 3.3.2 | Gaseous packaging methods

Packaging might not be considered a processing method, as processing often is thought of as something that aims to alter the product, whereas traditionally, packaging is mostly applied to maintain the product as it is. However, development within the field of food packaging has made it just as important as any other processing, especially within the field of mildly processed foods. Particular focus has been on MA packaging and active packaging (Yildirim et al., 2018).

The term modified atmosphere is often perceived as a synonym for modified atmosphere packaging, a postprocessing packaging step in which a mixture of different gases instantly replaces the air within a package at the time of sealing (Stammen et al., 1990). In reality, the term is much broader. Multiple methods exist for modifying the atmosphere within food packages, including vacuum packaging, emitters, SGS, and of course, MA packaging. Another storage principle is the controlled atmosphere storage (CAS), aiming to obtain the initial atmosphere in the storage system through storage (Yahia et al., 2019). CAS is widely used in the packaging of fruit and vegetables (Mditshwa et al., 2018; de Sigueira Oliveira et al., 2020) but is less used for seafood. However, an industrial application of SGS will need a prestep of CAS to obtain stable conditions before repacking in either MA or vacuum. Most of the SGS research has been performed in lab-scale experiments using the MAP principle to dissolve CO<sub>2</sub> into the product. However, an ongoing project funded by the Research Council of Norway (RCN) aims to develop a full-scale SGS technology concept for seafood (RCN, project number 294641).

The functional principle is the same regardless of the method chosen: the headspace within food packages is

altered to remove unwanted gases or introduce wanted ones. The main gases of importance are O<sub>2</sub>, N<sub>2</sub>, and CO<sub>2</sub>, whereas other constituents have been investigated and used (Sivertsvik et al., 2002). For most fish and seafood products, packaging aims at eliminating the presence of  $O_2$ , as  $O_2$  in most cases has deleterious effects on the quality of stored seafood products (Bouletis et al., 2017; Korpf & Mancini, 2014). As mentioned, bacterial spoilage of fish and seafood is often ascribed to the presence and growth of aerobic or facultative anaerobic bacteria (Gram & Huss, 1996) or lipid oxidation and development of rancidity (Mariutti & Bragagnolo, 2017). Hence, elimination of O<sub>2</sub> will provide extended shelf life by slowing bacterial spoilage and lipid oxidation. On the other hand, species with high trimethylamine oxide (TMAO) content often suffer from the removal of O2, as O2-depleted bacteria degrades TMAO to trimethylamine, causing formation of the characteristic "fishy" odor of spoiled seafood (Ashie et al., 1996). The removal of O<sub>2</sub> is the main purpose of vacuum packaging. Vacuum packaging was the first commercially developed MA packaging method. It consisted of packaging in low O<sub>2</sub> permeable materials after the evacuation of air, which under good vacuum conditions should reduce the O<sub>2</sub> concentration below 1% (DeWitt & Oliveira, 2016). Alternatively, flushing with  $N_2$  is used to replace  $O_2$  in packages as a measure for delaying spoilage. However, for the most part, N<sub>2</sub> is only used as a filler in MA packaging gas mixtures due to its solubility properties (Church & Parsons, 1995).

The most common application of gaseous packaging of fish and seafood is MA packaging. However, experimental findings vary significantly; mostly an extension in the range of 30%–60% of shelf life for fresh seafood is obtained when using an atmosphere with elevated CO<sub>2</sub> levels (Sivertsvik et al., 2002).

The use of high CO<sub>2</sub> in packaging headspace can change the composition of the microbiota of the product by favoring anaerobic or facultatively anaerobic species (Koutsoumanis et al., 2000; Noseda et al., 2012; Silbande et al., 2016). Brochothrix thermospacta is a potent spoilage bacterium that is considered the predominant spoilage organism of MA-packaged seafood products. Abel, Rotabakk, and Lerfall (2019) found MA packaging to reduce the growth of B. thermospacta compared to vacuum packaging. This agrees with Noseda et al.'s (2012) findings for pangasius fillets (Pangasius hypophthalmus). Rotabakk et al. (2008) found B. thermospacta not to be affected by increased CO<sub>2</sub> concentration. Moreover, Parlapani et al. (2014) reported MA packaging to favor the growth of B. thermospacta due to reduced competition. Differences in initial gas mixtures, g/p-ratio, or product characteristics could all be part of explaining these differences, although the exact reason is unknown. This shows the difficulty in comparing studies and hence estimating the effect of MA packaging (Sivertsvik et al., 2002).

Reduced-O<sub>2</sub> atmosphere may inhibit the growth of aerobic spoilage bacteria, but the same environment may be beneficial for strict or facultatively anaerobes such as Lis*teria* spp. High CO<sub>2</sub> levels have been shown to reduce the growth of Listeria spp. when used in MA packaging (Mejlholm & Dalgaard, 2007) by prolonging the lag phase as well as the growth rate (Abel, Rotabakk, & Lerfall, 2019; Pothuri et al., 1996; Provincial et al., 2013; Rutherford et al., 2007). However, it is essential to point out that these studies found MA packaging to cause a delay or slowdown of Listeria growth, not a complete inhibition. This is underlined by the predictive model developed by Mejlholm and Dalgaard (2007) showing that even an already preserved product as cold smoked Atlantic salmon needed a near 100% equilibrium dissolved CO<sub>2</sub> concentration to prevent the growth of L. monocytogenes.

Another concern regarding *L. monocytogenes* is the ability to form biofilms, a complex community of microorganisms attached to surfaces of food processing equipment. It has been shown that *L. monocytogenes* can form bacterial biofilms to survive on food processing surfaces under anaerobic conditions (Qian et al., 2020). Qian et al. (2020) investigated the influence of MA on *L. monocytogenes* biofilms during storage. They found that anaerobiosis significantly reduced the prevalence and thickness of biofilms formed compared to aerobic conditions.

Multiple methods have been used to evaluate the product quality of MA-packaged seafood. One of these is the evaluation of WHC or DL. It has been reported that increased CO<sub>2</sub> amount will alter WHC and hence increase DL (Davis, 1998; Masniyom et al., 2002; Randell et al., 1999; Rotabakk et al., 2008). However, when using SGS pretreatment or addition of CO<sub>2</sub> emitters in combination with MA packaging, no effect (Abel, Rotabakk, Rustad, et al., 2019) or even reduced DL (Hansen et al., 2007, 2009; Hansen, Mørkøre, Rudi, Rødbotten, et al., 2009; Rotabakk et al., 2008; Sivertsvik & Birkeland, 2006) was seen. A suggested explanation is an effect on the reduction in headspace volume, which occurs due to solubilization of CO<sub>2</sub> (Rotabakk et al., 2008).

Bouletis et al. (2014) evaluated color, appearance, odor, structure, flavor, and overall impression of squid on a fivegrade scale and found high  $CO_2$  content to increase the period with quality above the acceptable limit. Similar findings were made in almost all studies reporting sensory evaluation, with a tendency toward higher  $CO_2$  concentration resulting in shelf life extension (Hansen et al., 2016; Nikzade et al., 2019; Provincial et al., 2010).

The optimal use of  $CO_2$  MA packaging is limited by the solubility of  $CO_2$  causing volume changes and potentially packaging collapse when applied in flexible or semirigid

packages. A suggested solution is the use of  $CO_2$  emitters. Although SGS and MA packaging rely on changing the atmosphere before sealing the packages,  $CO_2$  emitters aim to obtain a CAS system by producing  $CO_2$  inside the packages through chemical reactions (Sivertsvik, 2003), hence classifying  $CO_2$  emitters as a form of active packaging (Yildirim et al., 2018). Other types of active packaging include oxygen scavengers, moisture scavengers, and antioxidant releaser (Yildirim et al., 2018).

Hansen et al. (2016) compared packaging of Atlantic cod in vacuum or MA packaging, with or without a CO<sub>2</sub> emitter, and found that inclusion of the emitter significantly decreased bacterial growth and prolonged sensory shelf life from 7 to 13 days. This agrees with Hansen et al. (2007), Hansen, Mørkøre, Rudi, Rødbotten, et al. (2009), and Tsironi et al. (2019). More importantly, the latter studies obtained increased shelf life, despite lowering the g/p ratio considerably, from 4:1 to 1.3:1 and 3:1 to 1:1, respectively (Hansen et al., 2007; Hansen, Mørkøre, Rudi, Rødbotten, et al., 2009). This means that less volume is needed for packaging, thus increasing transport and storage efficiency. Hansen et al. (2007, 2009) reported bulging of the top web as a result of the increased amount of CO<sub>2</sub> produced by the CO<sub>2</sub> emitter, which is one of the main disadvantages of emitter use. Relying on a chemical reaction within the packages makes it hard to control the CO<sub>2</sub> volume, especially as the facilitator, DL, varies significantly from sample to sample even within the same species.

#### 3.3.3 | Superchilling

A different approach to the mild inhibition methods and one of the most widely applied hurdles in the food industry is cold storage. In the industrial world, it has become so common that most of the time, it is not even considered an option but a necessity. Lowering the storage temperature has proven one of the most critical parameters affecting the growth of microorganisms. Similarly, reduced product temperature tends to slow enzymatic and other biochemical deterioration: the lower the temperature, the slower the deterioration (Kaale et al., 2011). The most used methods for fish and seafood are refrigerated ice storage (usually  $0-4^{\circ}C$ ) or frozen storage (-18 to  $-40^{\circ}C$ ) (Gallart-Jornet et al., 2007). Conventional freezing is often not preferred for high-quality products, as it may induce undesirable changes such as protein denaturation, reduced WHC, and increased DL on thawing (Kaale et al., 2011). The detrimental effect of freezing is triggered by the slow temperature decrease of conventional freezers, causing large ice crystals to form within the product. Large intracellular and extracellular ice crystals lead to rupture of cellular membranes and denaturation of cell components (Kaale & Eikevik, 2016; Kaale et al., 2011; Magnussen et al., 2008). Superchilling may act as an attractive compromise between conventional chilling and freezing (Duun & Rustad, 2008).

Superchilling, also known as partial freezing, implies temperatures in the borderline between chilling and freezing. During superchilling, the temperature of the food product is lowered  $1-2^{\circ}C$  below the initial freezing point of the product. This causes partial freezing of the product's water content (Kaale et al., 2011). The conversion of water into ice makes it less available for deteriorative processes. Furthermore, superchilling temperatures reduce microbial activity and prevent the growth of most bacteria (Kaale et al., 2011).

Use of superchilling has been reported for multiple fish and seafood products, including Atlantic salmon (*Salmo salar* L.) (Fernández et al., 2009; Sivertsvik et al., 2003), Atlantic cod (Eliasson et al., 2019; Sørensen et al., 2020), and crustaceans (Sun et al., 2017; Zeng et al., 2005). The general conclusion has been that superchilling results in an extension of the shelf life of stored food by 1.5 to four times compared to conventional chilling (Kaale et al., 2011; Magnussen et al., 2008).

The formation of ice crystals within the product can cause microstructural changes to the food. One potential effect of these changes is increased DL (Kaale et al., 2011), as reported by Bahuaud et al. (2008) and Liu et al. (2013). On the other hand, Sivertsvik et al. (2003) found no significant increase in DL, nor any other adverse effects on tested quality traits of superchilled Atlantic salmon. Similar reports were made by Duy Bao et al. (2007) for Arctic charr (Salvelinus alpinus). Chan et al. (2020), Cropotova et al. (2019), and Duun and Rustad (2007) even reported superchilling to decrease DL compared to traditional chilled or iced storage. Other potential negative effects of superchilling include degradation of texture and adverse sensory changes, as is reviewed by Banerjee and Maheswarappa (2019), Kaale et al. (2011), Magnussen et al. (2008), and Wu et al. (2014).

A downside to superchilling is the difficulty in maintaining appropriate storage temperatures with traditional consumer equipment such as refrigerators and freezers. A stable temperature is necessary to prevent ice melting and recrystallization, which may result in quality deterioration. These issues are easily overcome with industrial equipment, making superchilling a highly relevant application for commercial use. Furthermore, the ice formed on the surface of the products during superchilling will act as an internal ice reservoir, eliminating the need for external ice during transportation or short period storage. Ice usually makes up 20%–30% of the transported weight for ice-storage seafood (Magnussen et al., 2008); thus, superchilling could significantly increase transportation efficiency (Kaale et al., 2011).

A novel technology to preserve food is acidic electrolyzed water (AEW) ice (Ekonomou & Boziaris, 2021). AEW is produced in an electrolyzed water generator using water-added sodium chloride (NaCl) and voltage and current values of 9-10 V and 8-10 A, respectively. For the preservation of seafood, the AEW is normally frozen before use, and the term AEW ice is often used. AWE ice has been shown beneficial to improve the quality, shelf life, and food safety of several kinds of seafood, including shrimps (Lin et al., 2013; Wang et al., 2014; Zhang et al., 2015; Zhao et al., 2018) and squid (Xuan et al., 2017). Wang et al. (2014) suggested a primary mechanism improving the quality and safety of shrimps. Compared to traditional tap water ice, they reported AEW ice to limit the pH change, shrinkage of muscle fibers, and bacterial growth, whereas no adverse effects were reported on sarcoplasmic proteins. Moreover, they reported cathepsin B and polyphenol oxidase to be partly inhibited by the treatment. The working mechanisms causing the improved quality are not fully understood. However, it is known that chlorine and reactive oxygen can affect cell membranes and cause oxygen damage to DNA (Liao et al., 2018) and that factors such as pH (Park et al., 2004) and the oxidation-reduction potential (Liao et al., 2007) could be of significance. Most likely, there is a combination of mentioned reactions that inhibit bacterial growth and prevent product quality deterioration (Ekonomou & Boziaris, 2021).

#### 4 | HURDLE TECHNOLOGY

When applied individually, most of the aforementioned technologies are not sufficient to ensure food safety and shelf life. Hence, two or more technologies are often combined, either simultaneously or sequentially (Leistner & Gorris, 1995). The utilization of this approach is known as hurdle technology, barrier technology, or combination technology. "Hurdle technology" visualizes those technologies, or hurdles, that individually are to week to hamper spoilage alone. However, it might slow it down and if combined in sufficient numbers and correct height, it will be efficient in inhibiting microbial growth (Ashie et al., 1996). Hurdle technology is often applied to reduce the severity of the individual hurdles, thus lowering the adverse effect on nutritional and/or sensory quality (Tsironi et al., 2020).

The application of multiple technologies in one operation can act in one of three ways: (1) additively, (2) synergistic, or (3) antagonistic (Leistner & Gorris, 1995; Raso & Barbosa-Cánovas, 2003). Ates et al. (2016) found combined HP processing and mild heat treatment and showed a synergistic effect on the inactivation of *L. monocytogenes* as they obtained 6.62-log CFU/g reduction in inoculated fish soup after treatment using 500 MPa at 44°C compared

	Mild e heat	1	et al. – ark 13)						(Continue)
	Microwave	1	Murashita et al. (2017); Park et al. (2013)	.It					
(blue)	Ohmic heating	I		<ul><li>7); Choi et a</li><li>(2011)</li></ul>					
acterial samples	t Cold plasma	1	1	Kim et al. (2017); Choi et al. Kim et al. (2011) (2019)	I				
od products (red), or b	UV and pulsed light Cold plasma	1	Gayan et al. (2012); Kaya et al. (2015); Walkling-Ribeiro et al. (2008); Bradley et al. (2012); Marquenie et al. (2003)	Maktabi et al. (2011)	I	Colejo et al. (2018); Feizollahi et al. (2021)			
cts (black), other foo	High-pressure processing	1	Ates et al. (2016); Okozaki et al. (2000); Reddy et al. (1999); Reineke et al. (2011)		Min et al. (2016); Park et al. (2013)	I	Monteiro et al. (2018); Monteiro et al. (2019)		
oinations for fish produ	Curing	Beaufort et al. (2009); Chan et al. (2020); Lauzon et al. (2009); Midelet-Bourdin et al. (2008)	Gedela et al. (2007)	I	I	1	Heir et al. (2019); Holck et al. (2018)	Ekonomou et al. (2020); Erkan et al. (2011); Gómez-Estaca et al. (2007); Gudbjørnsdottir et al. (2010); Montero et al. (2007)	
rrdle technology coml	PEF	1	Aouir et al. (2015)	I	I	1	1	Gudmundsson & Hafsteinsson (2001)	
Examples of researched hurdle technology combinations for fish products (black), other food products (red), or bacterial samples (blue)	Modified atmosphere	Hansen et al. (2009); Sivertsvik et al. (2003); Sørensen et al. (2020); Wang et al. (2008)	Abel, Rotabakk, and Lerfall (2019); Abel, Rotabakk, Rustad, et al. (2019); Mohan et al. (2017)	Lerfall et al. (2018)	I	Olatunde et al. (2020); Yadav et al. (2020)	Monteiro et al. (2020); Rodrigues et al. (2016); Hinojosa et al. (2015)	Amanatidou et al. (2000); López-Caballero, Pérez-Mateos, Bonderías, et al. (2000); Rode et al. (2015)	
TABLE 1 Exa		Superchilling	Mild heat/sous vide	Microwave	Ohmic heating	Cold plasma	UV and pulsed light	High-pressure processing	

Mild e heat		
Microwave		
Ohmic heating		
Cold plasma		
UV and pulsed light Cold plasma		
High-pressure processing		
50		
Curing	e (I	
PEF	Cropotova, Tappi, Genovese, Rocculi, Laghi, et al. (2021); Cropotova, Tappi, Genovese, Rocculi, Dalla Rosa, et al. (2021)	
Modified atmosphere ]	Goktepe and Moody (1998); Mejlholm and Dalgaard (2007); Muratore and Licciardello (2005); Nilsson et al. (1997); Tsogas et al. (2019)	Chotphruethipong et al. (2019)
	Curing	PEF

(Continued)

**TABLE 1** 

to 2.64-log CFU/g reduction when using same pressure but lower (16°C) temperature, both for 5 min. A similar synergistic effect was reported for combined HP processing and MA packaging of Atlantic salmon (Amanatidou et al., 2000). On the contrary, Heinz and Knorr (2000) demonstrated that the lethality of PEF treatment applied in combination with sublethal HP processing on *Bacillus subtilis* vegetative cells was lower than the lethality of PEF treatment applied alone, thus indicating that HP treatment had a stabilizing effect.

Furthermore, combining processing technologies may facilitate the use of technologies that alone have been found infeasible for seafood, as is the case for ultrasound treatment. With ultrasonication, long processing times are needed to achieve sufficient bacterial inactivation, often resulting in adverse effects on the product (Lee et al., 2009; Sango et al., 2014). However, when combined with heat treatment (<53°C), a process known as thermoultrasonication, it was found to be an efficient processing method for shrimps, increasing the bacterial reduction from 0.6 to up to 4 log CFU/g, as well as reducing the processing time needed (Wang et al., 2013).

MA or other forms of CO<sub>2</sub> are often one of the most used methods in combinations. On the other hand, combinations of SGS with other processing methods are relatively new and have recently been reviewed by Esmaeilian et al. (2021). Rode et al. (2015) showed that use of SGS before HP processing reduced the average amount of L. innocua in fish soup by 1.5 log CFU/g, resulting in significantly lower content during storage. Furthermore, Abel, Rotabakk, and Lerfall (2019) showed pretreatment with SGS to cause a significant increase in heat inactivation of B. thermospacta compared to sous vide treated samples without SGS. Abel, Rotabakk, Rustad, et al. (2019) investigated the influence of a low-temperature treatment (40, 50, or 60°C) combined with various packaging technologies (MA or SGS) on both the microbial growth and the product quality in general. The study reported improved inhibition of L. innocua and highlighted the potential to obtain safe products even at low temperature sous vide treatments such as 40 and 50°C. This is in agreement with findings from milk samples (Loss & Hotchkiss, 2002). Combining MA packaging with superchilling increased shelf life of Atlantic cod from 13 days on ice storage to more than 32 days superchilled in MA packaging (13, 17, 23, and <32 days for iced air, iced MA, superchilled air, and superchilled MA, respectively), highlighting the synergistic nature of the hurdle combination (Sørensen et al., 2020).

An overview of some of the researched hurdle technology combinations in fish and seafood is presented in Table 1. When no research was found regarding fish or seafood, research on other food products or bacterial cultures was included. The addition of natural or artificial



preservatives or competitive microbiota is another hurdle often used in combinations. The table shows that multiple knowledge gaps exist regarding combination processing, especially for fish and seafood products. Some combinations are to be considered infeasible solely based on their different working mechanisms. For instance, CP relies on O<sub>2</sub> to produce ozone and other reactive species, which in turn causes the decontamination effect (Patil et al., 2014). On the other hand, MA packaging and SGS rely on the presence of CO<sub>2</sub> and the removal of O<sub>2</sub> to inhibit bacterial growth (Sivertsvik, 2000). Thus, a combination of these methods will probably prove inefficient if applied simultaneously. Other combinations have been proven efficient for other product types, such as pressure-ohmic thawing of beef and meat products (Min et al., 2016), but have not been tested for fish or seafood products.

Application of pressure-ohmic thawing showed a significantly improved quality of the meat compared to other thawing methods, based on texture, DL, and color, and significantly reduced thawing time (Min et al., 2016). Traditional freezing and thawing of seafood products are linked to quality deterioration. Hence, applying pressure-ohmic thawing could help improve the quality and offer an opportunity for long-distance transportation or storage.

Carbon dioxide to limit microbial growth is widely used, and Table 1 shows that effort has been taken to combine the bacteriostatic effect of  $CO_2$  with other mild technologies (Abel, Rotabakk, & Lerfall, 2019; Abel, Rotabakk, Rustad, et al., 2019; Lerfall et al., 2018; Rode et al., 2015). The combination of SGS with other technologies was recently reviewed by Esmaeilian et al. (2021). One of the key findings was that combining dissolved  $CO_2$  with other preservation technologies, as a hurdle technology, considerably enhanced the bacteriostatic effect of the treatments, mostly without compromising the product quality. Moreover, an industrial full-scale SGS concept is under development and will hopefully be available for the industry within a few years (Esmaeilian et al., 2021).

Other trending technologies in the food sector are PEF (Ekonomou & Boziaris, 2021) and HP processing (Ekonomou et al., 2020; Ekonomou & Boziaris, 2021; Shynkaryk et al., 2020). HP processing shows a comparative microbial inactivation to thermal processing, especially at higher pressures. A limitation is that too high pressures reduce the products' sensory and nutritional quality (Esmaeilian et al., 2021). However, as part of a hurdle approach, for example, in combination with  $CO_2$ , a considerable potential for the seafood industry exists. The combined effect of PEF with other technologies is attractive due to its potential to inhibit microorganisms and alter structural properties. Table 1 shows several gaps of potential combinations of PEF with other thermal

and nonthermal technologies of high interest for further research and industrial applications.

#### 5 | CONCLUSIVE REMARKS AND FURTHER OPPORTUNITIES

The general consumer's demand for minimally processed food is accompanied by shelf life and food safety challenges. Many mild processing technologies based on a variety of different working mechanisms have been developed to meet these challenges. This review on the current status of mild processing technologies and their effect on seafood safety, shelf life, and sensory quality shows that only a few of these technologies have been thoroughly studied and optimized for seafood products. Even fewer have been commercialized in the seafood processing industry. The reason is complex and consists of factors ranging from health, safety, and environmental issues on the processing plant (e.g., SGS) to high operating costs (HP processing), poor penetration depth (PL and UV), challenges due to unwanted color changes (HP processing), and increased product rancidity (CP). However, these challenges could be solved by developing novel technological concepts and/or using mild processing conditions in combination (Table 1). Most mild processing technologies are not sufficient alone to ensure food safety and proper shelf life. Hence more technologies are often combined to meet consumer's expectations and the food safety criteria set by national and international food authorities.

The present review has created an insight into and an overview of combinations of mild processing technologies with potential applications in the seafood industry. However, just as important as showing what is known, it visualizes the knowledge gaps and the potential for new technology developments in the sector (Table 1). Some methods, or combinations thereof, have shown great potential for nonseafood products, yet data are missing for fish and seafood. Table 1 highlights several of these gaps, which include, for example, the combination of mild heating (e.g., sous vide or MW) with more novel technologies such as PEF, PL, SGS, CP, or OH. However, before industrial applications are possible, more research is needed.

With the consumers' growing interest in mildly processed, fresh, and natural seafood options, there could be knowledge and profit from exploring these gaps, both for the academic sector and the industry.

#### ACKNOWLEDGMENT

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

#### AUTHOR CONTRIBUTIONS

Nanna Abel performed investigation and wrote the original draft. Bjørn Tore Rotabakk and Jørgen Lerfall supervised the study, and were responsible for editing the manuscript through the first and second revision. All authors conceptualized the idea of the study and reviewed and edited the original manuscript.

#### CONFLICT OF INTEREST

The authors declare no conflict of interest.

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#### REFERENCES

- Abel, N., Rotabakk, B. T., & Lerfall, J. (2019). Effect of heat treatment and packaging technology on the microbial load of lightly processed seafood. *LWT - Food Science and Technology*, 101, 123–129. https://doi.org/10.1016/j.lwt.2018.11.025
- Abel, N., Rotabakk, B. T., Rustad, T., Ahlsen, V. B., & Lerfall, J. (2019). Physiochemical and microbiological quality of lightly processed salmon (*Salmo salar L.*) stored under modified atmosphere. *Journal of Food Science*, 84(12), 3364–3372. https://doi.org/10.1111/ 1750-3841.14852
- Abel, N., Rotabakk, B. T., Rustad, T., & Lerfall, J. (2018). The influence of lipid composition, storage temperature, and modified atmospheric gas combinations on the solubility of CO<sub>2</sub> in a seafood model product. *Journal of Food Engineering*, 216, 151–158. https: //doi.org/10.1016/j.jfoodeng.2017.08.020
- Ahern, M., Thilsted, S. H., Oenema, S., Barange, M., Cartmill, M. K., Brandstrup, S. C., Doumeizel, V., Dyer, N., Frøyland, L., Garrido-Gamarro, E., Kühnhold, H., Mohammed, E., Penarubia, O., Potin, P., Sharan, S., Utheim, A., Uyar, B., Vannuccini, S., Ward, A., & Zhou, X. (2021). *The role of aquatic foods in sustainable healthy diets.* UN Nutrition.
- Al-Nehlawi, A., Saldo, J., Vega, L. F., & Guri, S. (2013). Effect of high carbon dioxide atmosphere packaging and soluble gas stabilization pre-treatment on the shelf-life and quality of chicken drumsticks. *Meat Science*, 94(1), 1–8. https://doi.org/10.1016/j.meatsci. 2012.12.008
- Albertos, I., Martín-Diana, A. B., Cullen, P. J., Tiwari, B. K., Ojha, S. K., Bourke, P., Álvarez, C., & Rico, D. (2017). Effects of dielectric barrier discharge (DBD) generated plasma on microbial reduction and quality parameters of fresh mackerel (*Scomber scombrus*) fillets. *Innovative Food Science and Emerging Technologies*, 44, 117–122. https://doi.org/10.1016/j.ifset.2017.07.006
- Albertos, I., Martin-Diana, A. B., Cullen, P. J., Tiwari, B. K., Ojha, S. K., Bourke, P., & Rico, D. (2019). Shelf-life extension of herring (*Clupea harengus*) using in-package atmospheric plasma technology. *Innovative Food Science and Emerging Technologies*, 53, 85–91. https://doi.org/10.1016/j.ifset.2017.09.010
- Allende, A., Tomás-Barberán, F. A., & Gil, M. I. (2006). Minimal processing for healthy traditional foods. *Trends in Food Science* and Technology, 17(9), 513–519. https://doi.org/10.1016/j.tifs.2006. 04.005

- Altintzoglou, T., Einarsdottir, G., Valsdottir, T., Schelvis, R., Skåra, T., & Luten, J. B. (2010). A voice-of-consumer approach in development of new seafood product concepts. *Journal of Aquatic Food Product Technology*, *19*(2), 130–145. https://doi.org/10.1080/ 10498850.2010.493636
- Altintzoglou, T., Hansen, K. B., Valsdottir, T., Odland, J. Ø., Martinsdóttir, E., Brunsø, K., & Luten, J. B. (2010). Translating barriers into potential improvements: The case of new healthy seafood product development. *Journal of Consumer Marketing*, 27(3), 224–235. https://doi.org/10.1108/07363761011038293
- Amanatidou, A., Schlüter, O., Lemkau, K., Gorris, L. G. M., Smid, E. J., & Knorr, D. (2000). Effect of combined application of high pressure treatment and modified atmospheres on the shelf life of fresh Atlantic salmon. *Innovative Food Science and Emerging Technologies*, 1(2), 87–98. https://doi.org/10.1016/S1466-8564(00)00007-2
- Aouir, A., Amiali, M., Kirilova-Gachovska, T., Benchabane, A., & Bitam, A. (2015). The effect of pulsed electric field (PEF) and ultrasoud (US) technologies on the extraction of phycopiliproteins from Arthrospira platensis. Paper presented at the 2015 IEEE Canada International Humanitarian Technology Conference (IHTC2015).
- Arason, S., van Nguyen, M., Thorarinsdottir, K. A., & Thorkelsson, G. (2014). Preservation of fish by curing. In I. S. Boziaris (Ed.), *Seafood processing, technology, quality and safety* (pp. 129–160). Wiley Blackwell.
- Arvanitoyannis, I. S., & Kotsanopoulos, K. V. (2013). Smoking of fish and seafood: History, methods and effects on physical, nutritional and microbiological properties. *Food and Bioprocess Technology*, 5(3), 831–853. https://doi.org/10.1007/s11947-011-0690-8
- Ashie, I. N. A., Smith, J. P., Simpson, B. K., & Haard, N. F. (1996). Spoilage and shelf-life extension of fresh fish and shellfish. *Critical Reviews in Food Science and Nutrition*, *36*(1–2), 87–121. https://doi.org/10.1080/10408399609527720
- Ates, M. B., Rode, T. M., Skipnes, D., & Lekang, O.-I. (2016). Modeling of *Listeria monocytogenes* inactivation by combined high-pressure and mild-temperature treatment in model soup. *European Food Research and Technology*, 242(2), 279–287. https://doi.org/10.1007/ s00217-015-2539-2
- Bahuaud, D., Mørkøre, T., Langsrud, Ø., Sinnes, K., Veiseth, E., Ofstad, R., & Thomassen, M. S. (2008). Effects of -1.5°C superchilling on quality of Atlantic salmon (*Salmo salar*) pre-rigor fillets: Cathepsin activity, muscle histology, texture and liquid leakage. *Food Chemistry*, 111(2), 329–339. https://doi.org/10.1016/ j.foodchem.2008.03.075
- Bak, K. H., Bolumar, T., Karlsson, A. H., Lindahl, G., & Orlien, V. (2019). Effect of high pressure treatment on the color of fresh and processed meats: A review. *Critical Reviews in Food Science and Nutrition*, 59(2), 228–252. https://doi.org/10.1080/10408398. 2017.1363712
- Baldwin, D. E. (2012). Sous vide cooking: A review. *International Journal of Gastronomy and Food Science*, 1(1), 15–30. https://doi.org/10.1016/j.ijgfs.2011.11.002
- Banerjee, R., & Maheswarappa, N. B. (2019). Superchilling of muscle foods: Potential alternative for chilling and freezing. *Critical Reviews in Food Science and Nutrition*, 59(8), 1256–1263. https://doi. org/10.1080/10408398.2017.1401975
- Banerjee, R., & Verma, A. K. (2015). Minimally processed meat and fish products. In M. W. Siddiqui & M. S. Rahman (Eds.), *Minimally*

processed foods - Technologies for safety, quality, and convenience (pp. 193–250). Springer International Publishing.

- Bank, H. L., John, J., Schmehl, M. K., & Dracht, R. J. (1990). Bactericidal effectiveness of modulated UV light. *Applied and Environmental Microbiology*, 58(12), 3888–3889. https://doi.org/10.1128/AEM. 56.12.3888-3889.1990
- Barba, F. J., Koubaa, M., do Prado-Silva, L., Orlien, V., & de Souza Sant'Ana, A. (2017). Mild processing applied to the inactivation of the main foodborne bacterial pathogens: A review. *Trends in Food Science and Technology*, 66, 20–35. https://doi.org/10.1016/j. tifs.2017.05.011
- Bastías, J. M., Moreno, J., Pia, C., Reyes, J., Quevedo, R., & Muñoz, O. (2015). Effect of ohmic heating on texture, microbial load, and cadmium and lead content of Chilean blue mussel (*Mytilus chilensis*). *Innovative Food Science and Emerging Technologies*, 30, 98– 102. https://doi.org/10.1016/j.ifset.2015.05.011
- Beaufort, A., Cardinal, M., Le-Bail, A., & Midelet-Bourdin, G. (2009). The effects of superchilled storage at -2°C on the microbiological and organoleptic properties of cold-smoked salmon before retail display. *International Journal of Refrigeration*, *32*(7), 1850– 1857. https://doi.org/10.1016/j.ijrefrig.2009.07.001
- Bintsis, T., Litopoulou-Tzanetaki, E., & Robinson, R. K. (2000). Existing and potential applications of ultraviolet light in the food industry - A critical review. *Journal of the Science of Food* and Agriculture, 80(6), 637–645. https://doi.org/10.1002/(SICI) 1097-0010(20000501)80:6(637::AID-JSFA603)3.0.CO;2-1
- Birkeland, S., Bencze Rørå, A. M., Skåra, T., & Bjerkeng, B. (2004). Effects of cold smoking procedures and raw material characteristics of product yield and quality parameters of cold smoked Atlantic salmon (*Salmo salar L.*) fillets. *Food Research International*, 37, 273–286.
- Birkeland, S., & Bjerkeng, B. (2005). The quality of cold-smoked Atlantic salmon (*Salmo salar*) as affected by salting method, time and temperature. *International Journal of Food Science and Technology*, 40(9), 963–976. https://doi.org/10.1111/j.1365-2621. 2005.01030.x
- Birkeland, S., & Rotabakk, B. T. (2014). Effects of additives and packaging method on quality and microbiological characteristics in mild thermal processed fish mince. *Journal of Aquatic Food Product Technology*, 23(4), 368–384. https://doi.org/10.1080/10498850. 2012.720004
- Birkeland, S., & Skåra, T. (2008). Cold smoking of Atlantic salmon (Salmo salar) fillet with smoke condensate - An alternative processing technology for the production of smoked salmon. Journal of Food Science, 73(6), s326–s332. https://doi.org/10.1111/j. 1750-3841.2008.00850.x
- Bjørnevik, M., Cardinal, M., Vallet, J.-L., Nicolaisen, O., & Arnarson, G. Ö. (2018). Effect of salting and cold-smoking procedure on Atlantic salmon originating from pre- or post rigor filleted raw material. Based on the measurement of physiochemical characteristics. *LWT - Food Science and Technology*, *91*, 431–438. https: //doi.org/10.1016/j.lwt.2018.01.047
- Bongiorno, T., Tulli, F., Comi, G., Sensidoni, A., Andyanto, D., & Iacumin, L. (2018). Sous vide cook-chill mussel (*Mytilus galloprovincialis*): Evaluation of chemical, microbiological and sensory quality during chilled storage (3°C). *LWT - Food Science and Technology*, *91*, 117–124. https://doi.org/10.1016/j.lwt.2017.12.005
- Bottino, F. O., Rodrigues, B. L., Ribeiro, J. D. N., Lázaro, C. A., & Conte, C. A., Jr. (2016). Effect of UV-C radiation on shelf-life of vacuum package Colossoma macropomum x Piaractus mesopotami-

cus fillets. Procedia Food Science, 7, 13–16. https://doi.org/10.1016/ j.profoo.2016.02.077

- Bouletis, A. D., Arvanitoyannis, I. S., & Hadjichristodoulou, C. (2017). Application of modified atmosphere packaging on aquacultured fish and fish products: A review. *Critical Reviews in Food Science* and Nutrition, 57(11), 2263–2285. https://doi.org/10.1080/10408398. 2013.862202
- Bouletis, A. D., Arvanitoyannis, I. S., Hadjichristodoulou, C., Neofitou, C., Sakkomitrou, M., & Kolokythopoulou, F. (2014). The effect of modified atmosphere packaging on the microbiological, physical, chemical and sensory characteristics of broadtail squid (*Illex coindetii*). *International Journal of Food Science and Technology*, 49(2), 329–336. https://doi.org/10.1111/ijfs.12286
- Boziaris, I. S. (2014). Seafood processing: Technology, quality and safety. John Wiley & Sons Inc.
- Bradley, D., McNeil, B., Laffey, J. G., & Rowan, N. J. (2012). Studies on the pathogenesis and survival of different culture forms of *Listeria monocytogenes* to pulsed UV-light irradiation after exposure to mild-food processing stresses. *Food Microbiology*, 30(2), 330–339. https://doi.org/10.1016/j.fm.2011.12.024
- Brody, A. L. (2012). The coming wave of microwave sterilization and pasteurization. *Food Technology Magazine*, 66(3), 78–80.
- Campus, M., Addis, M. F., Cappuccinelli, R., Porcu, M. C., Pretti, L., Tedde, V., Secchi, N., Stara, G., & Roggio, T. (2010). Stress relaxation behaviour and structural changes of muscle tissues from Gilthead sea bream (*Sparus aurata L.*) following high pressure treatment. *Journal of Food Engineering*, 96(2), 192–198. https://doi. org/10.1016/j.jfoodeng.2009.07.013
- Cardinal, M., Knockaert, C., Torrissen, O., Sigurgisladottir, S., Mørkøre, T., Thomassen, M., & Vallet, J. L. (2001). Relation of smoking parameters to the yield, colour and sensory quality of smoked Atlantic salmon (*Salmo salar*). *Food Research International*, 34(6), 537–550. https://doi.org/10.1016/S0963-9969(01) 00069-2
- Carlucci, D., Nocella, G., de Devitiis, B., Viscecchia, R., Bimbo, F., & Nardone, G. (2015). Consumer purchasing behaviour towards fish and seafood products. Patterns and insights from a sample of international studies. *Appetite*, *84*, 212–227. https://doi.org/10. 1016/j.appet.2014.10.008
- Casini, L., Contini, C., Romano, C., & Scozzafava, G. (2015). Trends in food consumptions: What is happening to generation *X*? *British Food Journal*, *117*(2), 705–718. https://doi.org/10.1108/ BFJ-10-2013-0283
- Chan, S. S., Roth, B., Skare, M., Hernar, M., Jessen, F., Løvdal, T., Jakobsen, A. N., & Lerfall, J. (2020). Effect of chilling technologies on water holding properties and other quality parameters throughout the whole value chain: From whole fish to cold-smoked fillets of Atlantic salmon (Salmo salar). Aquaculture, 526, 735381. https://doi.org/10.1016/j.aquaculture.2020.735381
- Chandrasekaran, S., Ramanathan, S., & Basak, T. (2013). Microwave food processing - A review. Food Research International, 52(1), 243–261. https://doi.org/10.1016/j.foodres.2013.02.033
- Cheigh, C.-I., Hwang, H.-J., & Chung, M.-S. (2013). Intense pulsed light (IPL) and UV-C treatments for inactivating *Listeria monocytogenes* on solid medium and seafoods. *Food Research International*, 54(1), 745–752. https://doi.org/10.1016/j.foodres.2013.08. 025
- Chéret, R., Chapleau, N., Delbarre-Ladrat, C., Verrez-Bagnis, V., & de Lamballerie, M. (2005). Effects of high pressure on texture and microstructure of sea bass (*Dicentrarchus labrax L.*) fillets.

Journal of Food Science, 70(8), e477–e483. https://doi.org/10.1111/ j.1365-2621.2005.tb11518.x

- Chiper, A. S., Chen, W., Mejlholm, O., Dalgaard, P., & Stamate, E. (2011). Atmospheric pressure plasma produced inside a closed package by a dielectric barrier discharge in Ar/CO<sub>2</sub> for bacterial inactivation of biological samples. *Plasma Sources Science and Technology*, *20*(2), 025008. https://doi.org/10.1088/0963-0252/20/2/025008
- Choi, M.-S., Cheigh, C.-I., Jeong, E.-A., Shin, J.-K., & Chung, M.-S. (2010). Nonthermal sterilization of *Listeria monocytogenes* in infant foods by intense pulsed-light treatment. *Journal of Food Engineering*, 97(4), 504–509. https://doi.org/10.1016/j.jfoodeng. 2009.11.008
- Choi, S., Puligundla, P., & Mok, C. (2016). Microbial decontamination of dried Alaska pollock shreds using corona discharge plasma jet: Effects on physicochemical and sensory characteristics. *Journal of Food Science*, *81*(4), M952–M957. https://doi.org/10.1111/1750-3841. 13261
- Choi, S., Puligundla, P., & Mok, C. (2017). Effect of corona discharge plasma on microbial decontamination of dried squid shreds including physico-chemical and sensory evaluation. *Food Science and Technology*, 75, 323–328. https://doi.org/10.1016/j.lwt. 2016.08.063
- Choi, W., Nguyen, L. T., Lee, S. H., & Jun, S. (2011). A microwave and ohmic combination heater for uniform heating of liquid-particle food mixtures. *Journal of Food Science*, 76(9), E576–E585. https:// doi.org/10.1111/j.1750-3841.2011.02413.x
- Chotphruethipong, L., Aluko, R. E., & Benjakul, S. (2019). Enhanced Asian sea bass skin defatting using porcine lipase with the aid of pulsed electric field pretreatment and vacuum impregnation. *Process Biochemistry*, 86, 58–64. https://doi.org/10.1016/j.procbio.2019. 08.012
- Christensen, L., Hovda, M. B., & Rode, T. M. (2017). Quality changes in high pressure processed cod, salmon and mackerel during storage. *Food Control*, 72, 90–96. https://doi.org/10.1016/j.foodcont. 2016.07.037
- Church, I. J., & Parsons, A. L. (1995). Modified atmosphere packaging technology: A review. *Journal of the Science of Food and Agriculture*, 67(2), 143–152. https://doi.org/10.1002/jsfa.2740670202
- Church, N. (1994). Developments in modified atmosphere packaging and related technologies. *Trends in Food Science and Technology*, *5*(11), 345–352. https://doi.org/10.1016/0924-2244(94)90211-9
- Colejo, S., Alvarez-Ordóñez, A., Prieto, M., González-Raurich, M., & López, M. (2018). Evaluation of ultraviolet light (UV), non-thermal atmospheric plasma (NTAP) and their combination for the control of foodborne pathogens in smoked salmon and their effect on quality attributes. *Innovative Food Science and Emerging Technologies*, 50, 84–93. https://doi.org/10.1016/j.ifset.2018.10.002
- Coyne, F. P. (1933). The effect of carbon dioxide on bacterial growth with special reference to the preservation of fish. Part II. *Journal of the Society of Chemical Industry*, *52*, 19–24.
- Creed, P. G. (1995). The sensory and nutritional quality of "sous vide" foods. *Food Control*, 6(1), 45–52. https://doi.org/10.1016/0956-7135(95)91453-R
- Critzer, F. J., Kelly-Wintenberg, K., South, S. L., & Golden, D. A. (2007). Atmospheric plasma inactivation of foodborne pathogens on fresh produce surfaces. *Journal of Food Protection*, 70(10), 2290– 2296. https://doi.org/10.4315/0362-028X-70.10.2290

- Cropotova, J., Mozuraityte, R., Standal, I. B., Grøvlen, M. S., & Rustad, T. (2019). Superchilled, chilled and frozen storage of Atlantic mackerel (*Scomber scombrus*) fillets - Changes in texture, drip loss, protein solubility and oxidation. *International Journal of Food Science and Technology*, 54(6), 2228–2235. https://doi.org/10.1111/ijfs. 14136
- Cropotova, J., Tappi, S., Genovese, J., Rocculi, P., Laghi, L., Dalla Rosa, M., & Rustad, T. (2021). Study of the influence of pulsed electric field pre-treatment on quality parameters of sea bass during brine salting. *Innovative Food Science & Emerging Technologies*, 70, 102706. https://doi.org/10.1016/j.ifset.2021.102706
- Cropotova, J., Tappi, S., Genovese, J., Rocculi, P., Dalla Rosa, M., & Rustad, T. (2021). The combined effect of pulsed electric field treatment and brine salting on changes in the oxidative stability of lipids and proteins and color characteristics of sea bass (*Dicentrarchus labrax*). *Heliyon*, 7(1), e05947. https://doi.org/10.1016/j. heliyon.2021.e05947
- Cruz-Romero, M., Smiddy, M., Hill, C., Kerry, J. P., & Kelly, A. L. (2004). Effects of high pressure treatment on physicochemical characteristics of fresh oysters (*Crassostrea gigas*). *Innovative Food Science and Emerging Technologies*, 5(2), 161–169. https://doi.org/ 10.1016/j.ifset.2004.01.002
- Daniels, J. A., Krishnamurthi, R., & Rizvi, S. S. H. (1985). A Review of effects of carbon dioxide on microbial growth and food quality. *Journal of Food Protection*, *48*(6), 532–537. https://doi.org/10.4315/0362-028X-48.6.532
- Davis, H. K. (1998). Fish and shellfish. In B. A. Blakistone (Ed.), Principles and applications of modified atmosphere packaging of foods (pp. 194–239). Blackie Academic and Professional.
- de Oliveira, F. A., Neto, O. C., dos Santos, L. M. R. d., Ferreira, E. H. R., & Rosenthal, A. (2017). Effect of high pressure on fish meat quality – A review. *Trends in Food Science & Technology*, 66, 1–19. https://doi.org/10.1016/j.tifs.2017.04.014
- de Oliveira, E. F., Nguyen, C. H., Stepanian, K., Cossu, A., & Nitin, N. (2019). Enhanced bacterial inactivation in apple juice by synergistic interactions between phenolic acids and mild food processing technologies. *Innovative Food Science and Emerging Technologies*, 56, 102186. https://doi.org/10.1016/j.ifset.2019.102186
- de Siqueira Oliveira, L., Eça, K. S., de Aquino, A. C., & da Silva, L. M. R. (2020). Modified and controlled atmosphere packaging. In M. W. Siddiqui (Ed.), *Fresh-cut fruits and vegetables* (pp. 151–164). Academic Press.
- de Souza Lira Santos, J., Mársico, E. T., Lemos, M., Cinquini, M. A., Alves da Silva, F., Dutra, Y. B., Franco, R. M., Conte, C. A., Jr., & Monteiro, M. L. G. (2018). Effect of the UV-C radiation on shelf life of vacuum-packed refrigerated pirarucu (*Arapaima gigas*) fillets. *Journal of Aquatic Food Product Technology*, 27(1), 48–60. https: //doi.org/10.1080/10498850.2017.1402840
- Del Giudice, T., Stranieri, S., Caracciolo, F., Ricci, E. C., Cembalo, L., Banterle, A., & Cicia, G. (2018). Corporate Social Responsibility certifications influence consumer preferences and seafood market price. *Journal of Cleaner Production*, 178, 526–533. https://doi.org/ 10.1016/j.jclepro.2017.12.276
- Devlieghere, F., Debevere, J., & Van Impe, J. (1998a). Concentration of carbon dioxide in the water-phase as a parameter to model the effect of a modified atmosphere on microorganisms. *International Journal of Food Microbiology*, *43*(1), 105–113. https://doi.org/ 10.1016/S0168-1605(98)00101-9



- Devlieghere, F., Debevere, J., & Van Impe, J. (1998b). Effect of dissolved carbon dioxide and temperature on the growth of *Lactobacillus sake* in modified atmospheres. *International Journal of Food Microbiology*, 41(3), 231–238. https://doi.org/10.1016/ S0168-1605(98)00052-X
- DeWitt, C., & Oliveira, A. (2016). Modified atmosphere systems and shelf life extension of fish and fishery products. *Foods*, *5*(3), 48. https://doi.org/10.3390/foods5030048
- Díaz, P., Garrido, M. D., & Bañón, S. (2011). Spoilage of sous vide cooked salmon (*Salmo salar*) stored under refrigeration. *Food Science and Technology International*, 17(1), 31–37. https://doi.org/10. 1177/1082013210368744
- Díaz, P., Nieto, G., Bañón, S., & Garrido, M. D. (2009). Determination of shelf life of sous vide salmon (*Salmo Salar*) based on sensory attributes. *Journal of Food Science*, 74(8), s371–s376. https: //doi.org/10.1111/j.1750-3841.2009.01317.x
- Duun, A. S., & Rustad, T. (2007). Quality changes during superchilled storage of cod (*Gadus morhua*) fillets. *Food Chemistry*, 105(3), 1067–1075. https://doi.org/10.1016/j.foodchem.2007.05.020
- Duun, A. S., & Rustad, T. (2008). Quality of superchilled vacuum packed Atlantic salmon (*Salmo salar*) fillets stored at -1.4 and -3.6°C. *Food Chemistry*, *106*(1), 122-131. https://doi.org/10.1016/j. foodchem.2007.05.051
- Duy Bao, H. N., Arason, S., & Thorarinsdottir, K. A. (2007). Effects of dry ice and superchilling on quality and shelf life of Arctic charr (*Salvelinus alpinus*) fillets. *International Journal of Food Engineering*, 3(3), 1–27. https://doi.org/10.2202/1556-3758.1093
- Ekonomou, S. I., Bulut, S., Karatzas, K. A. G., & Boziaris, I. S. (2020). Inactivation of *Listeria monocytogenes* in raw and hot smoked trout fillets by high hydrostatic pressure processing combined with liquid smoke and freezing. *Innovative Food Science and Emerging Technologies*, 64, 102427. https://doi.org/10.1016/j.ifset.2020. 102427
- Ekonomou, S. I., & Boziaris, I. S. (2021). Non-thermal methods for ensuring the microbiological quality and safety of seafood. *Applied Sciences*, 11(2), 833. https://doi.org/10.3390/app11020833
- Eliasson, S., Arason, S., Margeirsson, B., Bergsson, A. B., & Palsson, O. P. (2019). The effects of superchilling on shelf-life and quality indicators of whole Atlantic cod and fillets. *LWT - Food Science and Technology*, 100, 426–434. https://doi.org/10.1016/j.lwt.2018.10.062
- Erkan, N., Üretener, G., & Alpas, H. (2010). Effect of high pressure (HP) on the quality and shelf life of red mullet (*Mullus surmelutus*). *Innovative Food Science and Emerging Technologies*, 11(2), 569–264. https://doi.org/10.1016/j.ifset.2010.01.001
- Erkan, N., Üretener, G., Alpas, H., Selçuk, A., Özden, Ö., & Buzrul, S. (2011). The effect of different high pressure conditions on the quality and shelf life of cold smoked fish. *Innovative Food Science* and Emerging Technologies, 12(2), 104–110. https://doi.org/10.1016/ j.ifset.2010.12.004
- Ersoy, B., & Özeren, A. (2009). The effect of cooking methods on mineral and vitamin contents of African catfish. *Food Chemistry*, 115(2), 419–422. https://doi.org/10.1016/j.foodchem.2008.12.018
- Esmaeilian, S., Rotabakk, B. T., Lerfall, J., Jakobsen, A. N., Abel, N., Sivertsvik, M., & Olsen, A. (2021). The use of soluble gas stabilization technology on food – A review. *Trends in Food Science & Technology*, *118*, 154–166. https://doi.org/10.1016/j.tifs.2021.09.015
- Eto, H., Ono, Y., Ogino, A., & Nagatsu, M. (2008). Low-temperature sterilization of wrapped materials using flexible sheet-type dielec-

tric barrier discharge. *Applied Physics Letters*, 93(22), 221502. https://doi.org/10.1063/1.3039808

- EUMOFA. (2019). The EU fish market. Author.
- Faisal, M., Gani, A., & Mulana, F. (2019). Preliminary assessment of the utilization of durian peel liquid smoke as a natural preservative for mackerel. *F1000Research*, *8*, 240. https://doi.org/10.12688/ f1000research.18095.6
- Farkas, D. F., & Hoover, D. G. (2000). High pressure processing. Journal of Food Science, 65(Supplement 8), 47–64. https://doi.org/10. 1111/j.1750-3841.2000.tb00618.x
- Farr, D. (1990). High pressure technology in the food industry. Trends in Food Science and Technology, 1, 14–16. https://doi.org/10.1016/ 0924-2244(90)90004-I
- Feizollahi, E., Arshad, M., Yadav, B., Ullah, A., & Roopesh, M. S. (2021). Degradation of deoxynivalenol by atmospheric-pressure cold plasma and sequential treatments with heat and UV light. *Food Engineering Reviews*, 13, 696–705. https://doi.org/10.1007/ s12393-020-09241-0
- Fernandez-Gutierrez, S. A., Pedrow, P. D., Pitts, M. J., & Powers, J. (2010). Cold atmospheric-pressure plasmas applied to active packaging of apples. *IEEE Transactions on Plasma Science*, 38(4), 957– 965. https://doi.org/10.1109/TPS.2010.2042078
- Fernández, K., Aspe, E., & Roeckel, M. (2009). Shelf-life extension on fillets of Atlantic Salmon (*Salmo salar*) using natural additives, superchilling and modified atmosphere packaging. *Food Control*, 20(11), 1036–1042. https://doi.org/10.1016/j.foodcont.2008.12.010
- Ferreira, M., Almeida, A., Delgadillo, I., Saraiva, J., & Cunha, Â. (2016). Susceptibility of *Listeria monocytogenes* to high pressure processing: A review. *Food Reviews International*, 32(4), 377–399. https://doi.org/10.1080/87559129.2015.1094816
- Franco, D., Munekata, P. E. S., Agregán, R., Bermúdez, R., López-Pedrouso, M., Pateiro, M., & Lorenzo, J. M. (2020). Application of pulsed electric fields for obtaining antioxidant extracts from fish residues. *Antioxidants*, 9(2), 90. https://doi.org/10.3390/ antiox9020090
- Gallart-Jornet, L., Rustad, T., Barat, J. M., Fito, P., & Escriche, I. (2007). Effect of superchilled storage on the freshness and salting behaviour of Atlantic salmon (*Salmo salar*) fillets. *Food Chemistry*, 103(4), 1268–1281. https://doi.org/10.1016/j.foodchem.2006.10.040
- Garber, L. L., Hyatt, E. M., & Starr, R. G. (2003). Measuring consumer response to food products. *Food Quality and Preference*, 14(1), 3–15. https://doi.org/10.1016/S0950-3293(02)00030-7
- Garcia, G. W., Genigeorgis, C., & Lindroth, S. (1987). Risk of growth and toxin production by *Clostridium botulinum* nonproteolytic types B, E, and F in salmon fillets stored under modified atmospheres at low and abused temperatures. *Journal of Food Protection*, 50(4), 330–336. https://doi.org/10.4315/0362-028X-50.4.330
- Gaýan, E., Serrano, M. J., Monfort, S., Álvarez, I., & Condón, S. (2012). Combining ultraviolet light and mild temperatures for the inactivation of *Escherichia coli* in orange juice. *Journal of Food Engineering*, *113*(4), 598–605. https://doi.org/10.1016/j.jfoodeng.2012.07.018
- Gedela, S., Gamble, R. K., Macwana, S., Escoubas, J. R., & Muriana, P. M. (2007). Effect of inhibitory extracts derived from liquid smoke combined with postprocess pasteurization for control of *Listeria monocytogenes* on ready-to-eat meats. *Journal of Food Protection*, 70(12), 2749–2756. https://doi.org/10.4315/0362-028X-70.12.2749
- Gill, C. O. (1988). The solubility of carbon dioxide in meat. *Meat Science*, 22(1), 65–71. https://doi.org/10.1016/0309-1740(88)90027-7

- Goktepe, I., & Moody, M. W. (1998). Effect of modified atmosphere packaging on the quality of smoked catfish. *Journal of Muscle Foods*, *9*(4), 375–389. https://doi.org/10.1111/j.1745-4573.1998. tb00742.x
- Gómez-Estaca, J., Gómez-Guillén, M. C., & Montero, P. (2007). High pressure effects on the quality and preservation of cold-smoked dolphinfish (*Coryphaena hippurus*) fillets. *Food Chemistry*, 102(4), 1250–1259. https://doi.org/10.1016/j.foodchem.2006.07.014
- Gómez-López, V. M., Devlieghere, F., Bonduelle, V., & Debevere, J. (2005). Factors affecting the inactivation of micro-organisms by intense light pulses. *Journal of Applied Microbiology*, 99(3), 460– 470. https://doi.org/10.1111/j.1365-2672.2005.02641.x
- Gómez-López, V. M., Ragaert, P., Debevere, J., & Devlieghere, F. (2007). Pulsed light for food decontamination: A review. *Trends* in Food Science and Technology, 18(9), 464–473. https://doi.org/10. 1016/j.tifs.2007.03.010
- Gómez, B., Munekata, P. E. S., Gavahian, M., Barba, F. J., Martí-Quijal, F. J., Bolumar, T., Campagnol, P. C. B., Tomasevic, I., & Lorenzo, J. M. (2019). Application of pulsed electric fields in meat and fish processing industries: An overview. *Food Research International*, *123*, 95–105. https://doi.org/10.1016/j.foodres.2019.04.047
- Gonzalea-Fandos, E., & Laorden, A. (2020). Sous vide technology. In O. Yesim (Ed.), *Innovative technologies in seafood processing* (pp. 263–278). CRC Press.
- González-Fandos, E., Garciá-Linares, M. C., Villarino-Rodrigez, A., Farciá-Arias, M. T., & García-Fernández, M. C. (2004). Evaluation of the microbiological safety and sensory quality of rainbow trout (*Oncorhynchus mykiss*) processed by the sous vide method. Food Microbiology, 21(1), 193–201. https://doi.org/10.1016/ S0740-0020(03)00053-4
- Gou, J., Lee, H.-Y., & Ahn, J. (2010). Effect of high pressure processing on the quality of squid (*Todarodes pacificus*) during refrigerated storage. *Food Chemistry*, 119(2), 471–476. https://doi.org/10.1016/j. foodchem.2009.06.042
- Goulas, A. E., & Kontominas, M. (2005). Effect of salting and smoking-method on the keeping quality of chub mackerel (*Scomber japonicus*): Biochemical and sensory attributes. *Food Chemistry*, 93(3), 511–520. https://doi.org/10.1016/j.foodchem. 2004.09.040
- Govzman, S., Looby, S., Wang, X., Butler, F., Gibney, E., & Timon, C. (2021). A systematic review of the determinants of seafood consumption. *British Journal of Nutrition*, 126(1), 66–80. https://doi. org/10.1017/S0007114520003773
- Gram, L., & Huss, H. H. (1996). Microbiological spoilage of fish and fish products. *International Journal of Food Microbiology*, *33*(1), 121–137. https://doi.org/10.1016/0168-1605(96)01134-8
- Gudbjørnsdottir, B., Jonsson, A., Hafsteinsson, H., & Heinz, V. (2010). Effect of high-pressure processing on *Listeria* spp. and on the textural and microstructural properties of cold smoked salmon. *LWT* - *Food Science and Technology*, *43*(2), 366–374. https://doi.org/10.1016/j.lwt.2009.08.015
- Gudmundsson, M., & Hafsteinsson, H. (2001). Effect of electric field pulses on microstructure of muscle foods and roes. *Trends in Food Science & Technology*, *12*(3), 122–128. https://doi.org/10.1016/ S0924-2244(01)00068-1
- Guilbaud, M., Chafsey, I., Pilet, M.-F., Leroi, F., Prévost, H., Hébraud, M., & Dousset, X. (2008). Response of *Listeria monocytogenes* to liquid smoke. *Journal of Applied Microbiology*, 104(6), 1744–1753. https://doi.org/10.1111/j.1365-2672.2008.03731.x

- Guillén, M. D., Sopelana, P., & Partearroyo, M. A. (2000). Determination of polycyclic aromatic hydrocarbons in commercial liquid smoke flavorings of different compositions by gas chromatography-mass spectrometry. *Journal of Agriculture and Food Chemistry*, 48(2), 126–131. https://doi.org/10.1021/jf9908998
- Guillou, S., & Membré, J.-M. (2019). Inactivation of Listeria monocytogenes, Staphylococcus aureus, and Salmonella enterica under high hydrostatic pressure: A quantitative analysis of existing literature data. Journal of Food Protection, 82(10), 1802–1814. https: //doi.org/10.4315/0362-028X.JFP-19-132
- Gulzar, S., & Benjakul, S. (2020). Impact of pulsed electric field pretreatment on yield and quality of lipid extracted from cephalothorax of Pacific white shrimp (*Litopenaeus vannamei*) by ultrasoundassisted process. *International Journal of Food Science & Technol*ogy, 55(2), 619–630. https://doi.org/10.1111/ijfs.14316
- Guo, J., Huang, K., & Wang, J. (2015). Bactericidal effect of various non-thermal plasma agents and the influence of experimental conditions in microbial inactivation: A review. *Food Control*, 50, 482–490. https://doi.org/10.1016/j.foodcont.2014.09 .037
- Hansen, A. Å., Moen, B., Rødbotten, M., Berget, I., & Pettersen, M. K. (2016). Effect of vacuum or modified atmosphere packaging (MAP) in combination with a CO<sub>2</sub> emitter on quality parameters of cod loins (*Gadus morhua*). Food Packaging and Shelf Life, 9, 29–37. https://doi.org/10.1016/j.fpsl.2016.05. 005
- Hansen, A. Å., Mørkøre, T., Rudi, K., Langsrud, Ø., & Eie, T. (2009). The combined effect of superchilling and modified atmosphere packaging using CO<sub>2</sub> emitter on quality during chilled storage of pre-rigor salmon fillets (*Salmo salar*). *Journal of the Science of Food and Agriculture*, 89(10), 1625–1633. https://doi.org/10.1002/ jsfa.3599
- Hansen, A. Å., Mørkøre, T., Rudi, K., Olsen, E., & Eie, T. (2007). Quality changes during refrigerated storage of MA-packaged pre-rigor fillets of farmed Atlantic cod (*Gadus morhua* L.) using traditional MAP, CO<sub>2</sub> emitter, and vacuum. *Journal of Food Science*, 72(9), M423–M430. https://doi.org/10.1111/j.1750-3841.2007.00561.x
- Hansen, A. Å., Mørkøre, T., Rudi, K., Rødbotten, M., Bjerke, F., & Eie, T. (2009). Quality changes of prerigor filleted Atlantic salmon (*Salmo salar* L.) packaged in modified atmosphere using CO<sub>2</sub> emitter, traditional MAP, and vacuum. *Journal of Food Science*, 74(6), M242–M249. https://doi.org/10.1111/j.1750-3841.2009.01233.x
- Hattula, T., Elfving, K., Mroueh, U.-M., & Luoma, T. (2001). Use of liquid smoke flavouring as alternative to traditional flue gas smoking of rainbow trout fillets (Oncorhynchus mykiss). *LWT - Food Science and Technology*, 34(8), 521–525. https://doi.org/10.1006/fstl. 2001.0794
- He, H., Adams, R. M., Farkas, D. F., & Morrissey, M. T. (2002). Use of high-pressure processing for oyster shucking and shelf-life extension. *Journal of Food Science*, 67(2), 640–645. https://doi.org/10. 1111/j.1365-2621.2002.tb10652.x
- He, G., Yin, Y., Yan, X., & Yu, Q. (2014). Optimisation extraction of chondroitin sulfate from fish bone by high intensity pulsed electric fields. *Food Chemistry*, *164*, 205–210. https://doi.org/10.1016/j. foodchem.2014.05.032
- Heinrich, V., Zunabovic, M., Varzakas, T., Bergmair, J., & Kneifel, W. (2016). Pulsed light treatment of different food types with a special focus on meat: A critical review. *Critical Reviews in Food Science and Nutrition*, *56*(4), 591–613. https://doi.org/10.1080/10408398. 2013.826174

- Heinz, V., & Knorr, D. (2000). Effect of pH, ethanol addition and high hydrostatic pressure on the inactivation of *Bacillus subtilis* by pulsed electric fields. *Innovative Food Science and Emerging Technologies*, 1(2), 151–159. https://doi.org/10.1016/S1466-8564(00) 00013-8
- Heir, E., Liland, K. H., Carlehög, M., & Holck, A. L. (2019). Reduction and inhibition of *Listeria monocytogenes* in cold-smoked salmon by Verdad N6, a buffered vinegar fermentate, and UV-C treatments. *International Journal of Food Microbiology*, 291, 48–58. https://doi.org/10.1016/j.ijfoodmicro.2018.10.026
- Hinojosa, A., Gatica, I., Bustamante, A., Cárdenas, D., & Escalone, V. (2015). Effect of the combined treatment of UV-C light and modified atmosphere packaging on the inactivation of *Escherichia coli* inoculated watercress. *Journal of Food Processing and Preservation*, 39(6), 1525–1533. https://doi.org/10.1111/jfpp.12378
- Hite, B. H. (1899). The effect of pressure in the preservation of milk. *West Virginia Agricultural Experimental Station Bulletin*, *58*, 15–35. https://doi.org/10.33915/agnic.58
- Holck, A., Liland, K. H., Carlehög, M., & Heir, E. (2018). Reductions of *Listeria monocytogenes* on cold-smoked and raw salmon fillets by UV-C and pulsed UV light. *Innovative Food Science and Emerging Technologies*, 50, 1–10. https://doi.org/10.1016/j.ifset.2018.10. 007
- Huang, Y.-W., Leung, C.-K., Harrison, M. A., & Gates, K. W. (1993). Fate of *Listeria monocytogenes* and *Aeromonas hydrophila* on catfish fillets cooked in a microwave oven. *Journal of Food Science*, 58(3), 519–521. https://doi.org/10.1111/j.1365-2621.1993.tb04314.x
- Hughes, B. H., Perkins, L. B., Yang, T. C., & Skonberg, D. I. (2016). Impact of post-rigor high pressure processing on the physicochemical and microbial shelf-life of cultured red abalone (*Haliotis rufescens*). Food Chemistry, 194(1), 487–494. https://doi.org/10. 1016/j.foodchem.2015.07.144
- Hurtado, J. L., Montero, P., & Borderías, A. J. (2000). Extension of shelf life of chilled hake (*Merluccius capensis*) by high pressure. *Food Science and Technology International*, 6(3), 243–249. https:// doi.org/10.1177/108201320000600307
- Huss, H. H., Ababouch, L. H., & Gram, L. (2003) Assessment and management of seafood safety and quality. FAO Fisheries Technical Paper, no. 444.
- Jami, M., Ghanbari, M., Zunabovic, M., Domig, K. J., & Kneifel, W. (2014). Listeria monocytogenes in aquatic food products - A review. Comprehensive Reviews in Food Science and Food Safety, 13(5), 798– 813. https://doi.org/10.1111/1541-4337.12092
- Jermann, C., Koutchma, T., Margas, E., Leadley, C., & Ros-Polski, V. (2015). Mapping trends in novel and emerging food processing technologies around the world. *Innovative Food Science and Emerging Technologies*, 31, 14–27. https://doi.org/10.1016/j.ifset. 2015.06.007
- Jin, Y., Jiang, C., Jiao, Y., Llave, Y., Fukuoka, M., & Sakai, N. (2020). Analysis of ohmic heating of yellowtail (*Seriola quinqueradiata*) fillets at high frequencies by 3D simulation—Effect of ohmic heating system (batch and pseudo-continuous), sample shape, and size. *Innovative Food Science & Emerging Technologies*, 66, 102482. https://doi.org/10.1016/j.ifset.2020.102482
- Johnston, D. E., Farmer, L., Dynes, C., & Rutherford, J. (2003). High pressure processing of mussels, oysters and prawns. Paper presented at the Pressure to Succeed - An Insight into High Pressure Food Processing, Ireland.
- Kamalakanth, C. K., Ginson, J., Bindu, J., Venkateswarlu, R., Das, S., Chauhan, O. P., & Gopal, T. K. S. (2011). Effect of high pressure on

K-value, microbial and sensory characteristics of yellowfin tuna (*Thunnus albacares*) chunks in EVOH films during chill storage. *Innovative Food Science and Emerging Technologies*, *12*(4), 451–455. https://doi.org/10.1016/j.ifset.2011.06.001

- Kaya, Z., Yıldız, S., & Ünlütürk, S. (2015). Effect of UV-C irradiation and heat treatment on the shelf life stability of a lemon-melon juice blend: Multivariate statistical approach. *Innovative Food Science and Emerging Technologies*, 29, 230–239. https://doi.org/10. 1016/j.ifset.2015.03.005
- Kim, J.-W., Puligundla, P., & Mok, C. (2015). Microbial decontamination of dried laver using corona discharge plasma jet (CDPJ). *Journal of Food Engineering*, 161, 24–32. https://doi.org/10.1016/j. jfoodeng.2015.03.034
- Kim, J. E., Choi, H.-S., Lee, D.-U., & Min, S. C. (2017). Effects of processing parameters on the inactivation of *Bacillus cereus* spores on red pepper (*Capsicum annum* L.) flakes by microwave-combined cold plasma treatment. *International Journal of Food Microbiology*, 263, 61–66. https://doi.org/10.1016/j.ijfoodmicro.2017.09.014
- Kim, J. E., Oh, Y. J., Song, A. Y., & Min, S. C. (2019). Preservation of red pepper flakes using microwave-combined cold plasma treatment. *Journal of the Science of Food and Agriculture*, 99(4), 1577– 1585. https://doi.org/10.1002/jsfa.9336
- Kjällstrand, J., & Petersson, G. (2001). Phenolic antioxidants in wood smoke. *The Science of the Total Environment*, 277(1), 69–75. https://doi.org/10.1016/S0048-9697(00)00863-9
- Knoche, W. (1980). Chemical reactions of CO<sub>2</sub> in water. In C. Bauer,
  G. Gros, & H. Bartels (Eds.), *Biophysics and physiology of carbon dioxide* (pp. 3–11). Springer.
- Korpf, D., & Mancini, M. (2014). Modified and controlled atmosphere. In M. Dikeman & C. Devine (Eds.), *Encyclopedia of meat sciences* (2nd ed., Vol. 3, pp. 9–12). Elsevier Academic Press.
- Kostyra, E., & Barłyko-Pikielna, N. (2006). Volatiles composition and flavour profile identity of smoke flavourings. *Food Quality and Preference*, *17*(1–2), 85–95. https://doi.org/10.1016/j.foodqual.2005. 06.008
- Koutsoumanis, K. P., Taoukis, P. S., Drosinos, E. H., & Nychas, G.-J. (2000). Applicability of an Arrhenius model for the combined effect of temperature and CO<sub>2</sub> packaging on the spoilage microflora of fish. *Applied and Environmental Microbiology*, 66(8), 3528–3534. https://doi.org/10.1128/AEM.66.8.3528-3534.2000
- Kramer, B., Wunderlich, J., & Muranyi, P. (2017). Recent findings in pulsed light disinfection. *Journal of Applied Microbiology*, 122(4), 830–856. https://doi.org/10.1111/jam.13389
- Kulawik, P., Alvarez, C., Cullen, P. J., Aznar-Roca, R., Mullen, A. M., & Tiwari, B. K. (2018). The effect of non-thermal plasma on the lipid oxidation and microbiological quality of sushi. *Innovative Food Science and Emerging Technologies*, 45, 412–417. https://doi.org/10.1016/j.ifset.2017.12.011
- Kulawik, P., & Tiwari, B. K. (2019). Recent advancements in the application of non-thermal plasma technology for the seafood industry. *Critical Reviews in Food Science and Nutrition*, *59*(19), 3199–3210. https://doi.org/10.1080/10408398.2018.1510827
- Kumar, T. (2018). A review on ohmic heating technology: Principle, applications and scope. *International Journal of Agriculture, Environment and Biotechnology*, *11*(4), 679–687. https://doi.org/10. 30954/0974-1712.08.2018.10
- Kaale, L. D., & Eikevik, T. M. (2016). Changes of proteins during superchilled storage of Atlantic salmon muscle (*Salmo salar*). *Journal of Food Science and Technology*, 53(1), 441–450. https://doi. org/10.1007/s13197-015-1979-9

- Kaale, L. D., Eikevik, T. M., Rustad, T., & Kolsaker, K. (2011). Superchilling of food: A review. *Journal of Food Engineering*, 107, 141– 146. https://doi.org/10.1016/j.jfoodeng.2011.06.004
- Lakshmanan, R., Parkinson, J. A., & Piggott, J. R. (2007). Highpressure processing and water-holding capacity of fresh and coldsmoked salmon (*Salmo salar*). *LWT - Food Science and Technology*, 40(3), 544–551. https://doi.org/10.1016/j.lwt.2005.12.003
- Lascorz, D., Torella, E., Lyng, J. G., & Arroyo, C. (2016). The potential of ohmic heating as an alternative to steam for heat processing shrimps. *Innovative Food Science and Emerging Technologies*, 37, 329–335. https://doi.org/10.1016/j.ifset.2016.06.014
- Lauzon, H. L., Magnússon, H., Sveinsdottir, K., Gudjónsdóttir, M., & Martinsdóttir, E. (2009). Effect of brining, modified atmosphere packaging, and superchilling on the shelf life of cod (*Gadus morhua*) loins. *Journal of Food Science*, 74(6), M258–M267. https: //doi.org/10.1111/j.1750-3841.2009.01200.x
- Lawley, M., Craig, J. F., Dean, D., & Birch, D. (2019), The role of seafood sustainability knowledge in seafood purchase decisions. *British Food Journal*, 121(10), 2337–2350. https://doi.org/10.1108/ BFJ-08-2018-0513
- Lee, E.-S., Park, S. Y., & Ha, S.-D. (2015). Effect of UV-C light on the microbial and sensory quality of seasoned dried seafood. *Food Sci*ence and Technology International, 22(3), 213–220. https://doi.org/ 10.1177/1082013215586293
- Lee, H., Zhou, B., Liang, W., Feng, H., & Martin, S. (2009). Inactivation of *Escherichia coli* cells with sonication, manosonication, thermosonication, and manothermosonication: Microbial responses and kinetics modeling. *Journal of Food Engineering*, 93(3), 354–364. https://doi.org/10.1016/j.jfoodeng.2009.01. 037
- Leistner, L., & Gorris, L. G. M. (1995). Food preservation by hurdle technology. *Trends in Food Science and Technology*, 6(2), 41–46. https://doi.org/10.1016/S0924-2244(00)88941-4
- Lerfall, J., Akse, L., Østerlie, M., & Birkeland, S. (2011). Salting method affects the retention of carotenoids in the fillet surface of cold-smoked Atlantic salmon (*Salmo salar L.*). *International Journal of Food Science and Technology*, 46(10), 2218–2223. https: //doi.org/10.1111/j.1365-2621.2011.02723.x
- Lerfall, J., Nordeng Jakobsen, A., Skipnes, D., Waldenstrøm, L., Hoel, S., & Rotabakk, B. T. (2018). Comparative evaluation of the quality and shelf life of Atlantic salmon (*Salmo salar* L.) filets using microwave and conventional pasteurization in combination with novel packaging methods. *Journal of Food Science*, 83(12), 3099– 3109. https://doi.org/10.1111/1750-3841.14384
- Leroi, F., Joffraud, J. J., Chevalier, F., & Cardinal, M. (1998). Study of the microbial ecology of cold-smoked salmon during storage at 8°C. *International Journal of Food Microbiology*, 39(1), 111–121. https://doi.org/10.1016/S0168-1605(97)00126-8
- Leroi, F., Joffraud, J. J., Chevalier, F., & Cardinal, M. (2001). Research of quality indices for cold-smoked salmon using a stepwise multiple regression of microbiological counts and physico-chemical parameters. *Journal of Applied Microbiology*, *90*(4), 578–587. https: //doi.org/10.1046/j.1365-2672.2001.01283.x
- Liao, L. B., Chen, W. M., & Xiao, X. M. (2007). The generation and inactivation mechanism of oxidation–reduction potential of electrolyzed oxidizing water. *Journal of Food Engineering*, 78(4), 1326– 1332. https://doi.org/10.1016/j.jfoodeng.2006.01.004
- Liao, X., Liu, D., Xiang, Q., Ahn, J., Chen, S., Ye, X., & Ding, T. (2017). Inactivation mechanisms of non-thermal plasma on microbes: A

review. Food Control, 75, 83–91. https://doi.org/10.1016/j.foodcont. 2016.12.021

- Liao, X., Su, Y., Liu, D., Chen, S., Hu, Y., Ye, X., Wang, J., & Ding, T. (2018). Application of atmospheric cold plasma-activated water (PAW) ice for preservation of shrimps (*Metapenaeus ensis*). *Food Control*, 94, 307–314. https://doi.org/10.1016/j.foodcont.2018. 07.026
- Lin, T., Wang, J. J., Li, J. B., Liao, C., Pan, Y. J., & Zhao, Y. (2013). Use of acidic electrolyzed water ice for preserving the quality of shrimp. *Journal of Agricultural and Food Chemistry*, *61*(36), 8695– 8702. https://doi.org/10.1021/jf4019933
- Linton, M., Mc Clements, J. M. J., & Patterson, M. F. (2003). Changes in the microbiological quality of shellfish, brought about by treatment with high hydrostatic pressure. *International Journal of Food Science and Technology*, *38*(6), 713–727. https://doi.org/10.1046/j. 1365-2621.2003.00724.x
- Liu, D., Liang, L., Xia, W., Regenstein, J. M., & Zhou, P. (2013). Biochemical and physical changes of grass carp (*Ctenopharyngodon idella*) fillets stored at -3 and 0°C. *Food Chemistry*, 140(1-2), 105– 114. https://doi.org/10.1016/j.foodchem.2013.02.034
- Liu, L., Llave, Y., Jin, Y., Zheng, D.-y., Fukuoka, M., & Sakai, N. (2017). Electrical conductivity and ohmic thawing of frozen tuna at high frequencies. *Journal of Food Engineering*, 197, 68–77. https://doi. org/10.1016/j.jfoodeng.2016.11.002
- Llave, Y., Morinaga, K., Fukuoka, M., & Sakai, N. (2018). Characterization of ohmic heating and sous-vide treatment of scallops: Analysis of electrical conductivity and the effect of thermal protein denaturation on quality attribute changes. *Innovative Food Science & Emerging Technologies*, 50, 112–123. https://doi.org/10. 1016/j.ifset.2018.09.007
- Loss, C. R., & Hotchkiss, J. (2002). Effect of dissolved carbon dioxide on thermal inactivation of microorganisms in milk. *Journal* of food protection, 65(12), 1924–1929. https://doi.org/10.4315/0362-028X-65.12.1924
- López-Caballero, M. E., Pérez-Mateos, M., Bonderías, A. J., & Montero, P. (2000). Extension of shelf life of prawns (*Penaeus japonicus*) by vacuum packaging and high-pressure treatment. *Journal of Food Protection*, 63(10), 1381–1388. https://doi.org/10.4315/ 0362-028X-63.10.1381
- López-Caballero, M. E., Pérez-Mateos, M., Montero, M. P., & Borderías, A. J. (2000). Oyster preservation by high-pressure treatment. *Journal of Food Protection*, 63(2), 196–201. https://doi.org/ 10.4315/0362-028X-63.2.196
- Løvdal, T. (2015). The microbiology of cold smoked salmon. Food Control, 54, 360–373. https://doi.org/10.1016/j.foodcont.2015.02. 025
- MacGregor, S. J., Rowan, N. J., McIlvaney, L., Anderson, J. G., Fouracre, R. A., & Farish, O. (1998). Light inactivation of foodrelated pathogenic bacteria using a pulsed power source. *Letters in Applied Microbiology*, 27(2), 67–70. https://doi.org/10.1046/ j.1472-765X.1998.00399.x
- Maesano, G., Carra, G., & Vindigni, G. (2019). Sustainable dimensions of seafood consumer purchasing behaviour: A review: Acces La Success. Calitatea, 20, 358–364. https://www.proquest.com/scholarly-journals/sustainable-dimensions-seafood-consumer/docview/2198414196/se-2?accountid=12870
- Magnussen, O. M., Haugland, A., Torstveit Hemmingsen, A. K., Johansen, S., & Nordtved, T. S. (2008). Advances in superchilling of food - Process characteristics and product quality. *Trends in Food*

*Science and Technology*, *19*, 418–424. https://doi.org/10.1016/j.tifs. 2008.04.005

- Makroo, H. A., NK, R., & Srivastava, B. (2020). Ohmic heating assisted inactivation of enzymes and microorganisms in foods: A review. *Trends in Food Science and Technology*, 97, 451–465. https: //doi.org/10.1016/j.tifs.2020.01.015
- Maktabi, S., Watson, I., & Parton, R. (2011). Synergistic effect of UV, laser and microwave radiation or conventional heating on *E. coli* and on some spoilage and pathogenic bacteria. *Innovative Food Science and Emerging Technologies*, 12(2), 129–134. https://doi.org/ 10.1016/j.ifset.2010.12.011
- Mañas, P., & Pagán, R. (2005). Microbial inactivation by new technologies of food preservation. *Journal of Applied Microbiology*, 98(6), 1387–1399. https://doi.org/10.1111/j.1365-2672.2005.02561.x
- Mandal, R., Mohammadi, X., Wiktor, A., Singh, A., & Pratap Singh, A. (2020). Applications of pulsed light decontamination technology in food processing: An overview. *Applied Sciences*, 10(10), 3606. https://doi.org/10.3390/app10103606
- Mariutti, L. R. B., & Bragagnolo, N. (2017). Influence of salt on lipid oxidation in meat and seafood products: A review. *Food Research International*, *94*, 90–100. https://doi.org/10.1016/j.foodres.2017. 02.003
- Marquenie, D., Geeraerd, A. H., Lammertyn, J., Soontjens, C., Van Impe, J. F., Michielis, C. W., & Nicolaï, B. M. (2003). Combinations of pulsed white light and UV-C or mild heat treatment to inactivate conidia of *Botrytis cinerea* and *Monilia fructigena*. *International Journal of Food Microbiology*, 85(1–2), 185–196. https://doi. org/10.1016/S0168-1605(02)00538-X
- Martínez-Monteagudo, S. I., & Balasubramaniam, V. M. (2016). Fundamentals and applications of high-pressure processing technology. In V. M. Balasubramaniam, G. V. Barbosa-Cánovas, & H. Lelieveld (Eds.), *High pressure processing of food: Principles, technology and applications* (pp. 3–17). Springer.
- Martinez, O., Salmerón, J., Guillén, M., & Casas, C. (2007). Textural and physicochemical changes in salmon (*Salmo salar*) treated with commercial liquid smoke flavourings. *Food Chemistry*, 100(2), 498–503. https://doi.org/10.1016/j.foodchem.2005.09. 071
- Martinez, O., Salmerón, J., Guillén, M. D., Pin, C., & Casas, C. (2012). Physicochemical, sensorial and textural characteristics of liquid smoked salmon (*Salmo salar*) as affected by salting treatment and sugar addition. *International Journal of Food Science and Technology*, 47(5), 1086–1096. https://doi.org/10.1111/j.1365-2621.2012. 02945.x
- Masniyom, P., Benjakul, S., & Visessanguan, W. (2002). Shelf-life extension of refrigerated seabass slices under modified atmosphere packaging. *Journal of the Science of Food and Agriculture*, 82(8), 873–880. https://doi.org/10.1002/jsfa.1108
- Mejlholm, O., & Dalgaard, P. (2007). Modeling and predicting the growth boundary of *Listeria monocytogenes* in lightly preserved seafood. *Journal of Food Protection*, 70(1), 70–84. https://doi.org/ 10.4315/0362-028X-70.1.70
- Mendes de Souza, P., Briviba, K., Müller, A., Fernández, A., & Stahl, M. (2013). Cyto-genotoxic and oxidative effects of a continuous UV-C treatment of liquid egg products. *Food Chemistry*, 138(2-3), 1682–1688. https://doi.org/10.1016/j.foodchem.2012.11.105
- Mendes, R., & Gonçalves, A. (2008a). Effect of soluble CO<sub>2</sub> stabilisation and vacuum packaging on the shelf life of farmed sea bream and sea bass fillets. *International Journal of Food Science and Tech*-

*nology*, *43*(9), 1678–1687. https://doi.org/10.1111/j.1365-2621.2008. 01737.x

- Mendes, R., & Gonçalves, A. (2008b). Effect of soluble CO<sub>2</sub> stabilization on the quality of fillets from farmed gilthead sea bream (*Sparus aurata*) and European sea bass (*Dicentrarchus labrax*). *Journal of Aquatic Food Product Technology*, 17(4), 342–366. https: //doi.org/10.1080/10498850802369187
- Mendes, R., Silva, H. A., Anacleto, P., & Cardoso, C. (2011). Effect of CO<sub>2</sub> dissolution on the shelf life of ready-to-eat Octopus vulgaris. Innovative Food Science and Emerging Technologies, 12(4), 551–561. https://doi.org/10.1016/j.ifset.2011.07.003
- Menozzi, D., Nguyen, T. T., Sogari, G., Taskov, D., Lucas, S., Castro-Rial, J. L. S., & Mora, C. (2020). Consumers' preferences and willingness to pay for fish products with health and environmental labels: Evidence from five European countries. *Nutrients*, 12(9), 2650. https://doi.org/10.3390/nu12092650
- Metaxas, A. C., & Meredith, R. J. (1988). *Industrial microwave heating*. Peter Peregrinus Ltd.
- Midelet-Bourdin, G., Beaufort, A., Leroi, F., Cardinal, M., Rudelle, S., Leleu, G., Copin, S., & Malle, P. (2008). Impact of -2 degrees C superchilling before refrigerated storage (4 and 8 degrees C) on the microbiological and sensory qualities of cold-smoked salmon. *Journal of Food Protection*, *71*(11), 2198. https://doi.org/10.4315/ 0362-028X-71.11.2198
- Min, S.-G., Hong, G.-P., Chun, J.-Y., & Park, S. H. (2016). Pressure ohmic thawing: A feasible approach for the rapid thawing of frozen meat and its effects on quality attributes. *Food and Bioprocess Technology*, 9(4), 564–575. https://doi.org/10.1007/s11947-015-1652-3
- Mditshwa, A., Fawole, O. A., & Opara, U. L. (2018). Recent developments on dynamic controlled atmosphere storage of apples—A review. Food Packaging and Shelf Life, 16, 59–68. https://doi.org/ 10.1016/j.fpsl.2018.01.011
- Misra, N., Segat, A., & Cullen, P. J. (2015). Atmospheric-pressure nonthermal plasma decontamination of food. In R. V. Ravishankar (Ed.), Advances in food biotechnology (pp. 565–574). John Wiley & Sons Ltd.
- Misra, N., Tiwari, B. K., Raghavarao, K. S. M. S., & Cullen, P. J. (2011). Nonthermal plasma inactivation of food-borne pathogens. *Food Engineering Reviews*, 3(3-4), 159–170. https://doi.org/10.1007/ s12393-011-9041-9
- Mohan, C. O., Ravishankar, C. N., & Srinivasa Gopal, T. K. (2017). Effect of vacuum packaging and sous vide processing on the quality of Indian white shrimp (*Fenneropenaeus indicus*) during chilled storage. *Journal of Aquatic Food Product Technology*, 26(10), 1280– 1292. https://doi.org/10.1080/10498850.2016.1236869
- Mol, S., Ozturan, S., & Cosansu, S. (2012). Determination of the quality and shelf life of sous vide packaged bonito (*Sarda sarda*, bloch 1793) stored at 4 and 12 C. *Journal of Food Quality*, *35*(2), 137–143. https://doi.org/10.1111/j.1745-4557.2011.00430.x
- Molina, B., Sáez, M. I., Martínez, T. F., Guil-Guerrero, J. L., & Suárez, M. D. (2014). Effect of ultraviolet light treatment on microbial contamination, some textural and organoleptic parameters of cultured sea bass fillets (*Dicentrarchus labrax*). *Innovative Food Science and Emerging Technologies*, 26, 205–213. https://doi.org/10.1016/j.ifset. 2014.07.002
- Montazeri, N., Himelbloom, B., Oliveira, A., Leigh, M. B., & Crapo, C. (2013). Refined liquid smoke: A potential antilisterial additive to cold-smoked sockeye salmon (*Oncorhynchus nerka*). Journal of

Food Protection, 76(5), 812–819. https://doi.org/10.4315/0362-028X. JFP-12-368

- Montazeri, N., Oliveira, A. C. M., Himelbloom, B. H., Leigh, M. B., & Crapo, C. A. (2013). Chemical characterization of commercial liquid smoke products. *Food Science and Nutrition*, 1(1), 102–115. https://doi.org/10.1002/fsn3.9
- Monteiro, M. L. G., Mársico, E. T., Canto, A. C. V., Costa-Lima, B. R. C., Costa, M. P., Viana, F. M., da Silva, T. J. P., & Conte, C. A., Jr. (2017). Impact of UV-C light on the fatty acid profile and oxidative stability of Nile tilapia (*Oreochromis niloticus*) fillets. *Journal of Food Science*, 82(4), 1028–1036. https://doi.org/10.1111/1750-3841. 13685
- Monteiro, M. L. G., Mársico, E. T., Mano, S. B., da Silveira Alvares, T., Rosenthal, A., Lemos, M., Ferrari, E., Lázaro, C. A., & Conte, C. A., Jr. (2018). Combined effect of high hydrostatic pressure and ultraviolet radiation on quality parameters of refrigerated vacuumpacked tilapia (*Oreochromis niloticus*) fillets. *Scientific Reports*, 8(1), 9524–9511. https://doi.org/10.1038/s41598-018-27861-9
- Monteiro, M. L. G., Mársico, E. T., Mutz, Y., Castro, V. S., Moreira, R. V., Álvares, T., & Conte, C. A., Jr. (2020). Combined effect of oxygen-scavenger packaging and UV-C radiation on shelf life of refrigerated tilapia (*Oreochromis niloticus*) fillets. *Scientific Reports*, 10(1), 4243–4212. https://doi.org/10.1038/ s41598-020-61293-8
- Monteiro, M. L. G., Mársico, E. T., Rosenthal, A., & Conte, C. A., Jr. (2019). Synergistic effect of ultraviolet radiation and high hydrostatic pressure on texture, color, and oxidative stability of refrigerated tilapia fillets. *Journal of the Science of Food and Agriculture*, 99(9), 4474–4481. https://doi.org/10.1002/jsfa.9685
- Montero, M. P., Gomez-Estaca, J., & Gomez-Guillen, M. C. (2007). Influence of salt, smoke, and high pressure on growth of *Listeria monocytogenes* and spoilage microflora on cold-smoked dolphinfish (*Coryphaena hippurus*). *Journal of Food Protection*, 70(2), 399– 404. https://doi.org/10.4315/0362-028X-70.2.399
- Murashita, S., Kawamura, S., & Koseki, S. (2017). Effects of ohmic heating, including electric field intensity and frequency, on thermal inactivation of Bacillus subtilis spores. *Journal of Food Protection*, 80(1), 164–168. https://doi.org/10.4315/0362-028X.JFP-16-300
- Muratore, G., & Licciardello, F. (2005). Effect of vacuum and modified atmosphere packaging on the shelf-life of liquid-smoked swordfish (*Xiphias gladius*) slices. *Journal of Food Science*, 70(5), C359– C363. https://doi.org/10.1111/j.1365-2621.2005.tb09967.x
- Murchie, L. W., Cruz-Romero, M., Kerry, J. P., Linton, M., Patterson, M. F., Smiddy, M., & Kelly, A. L. (2005). High pressure processing of shellfish: A review of microbiological and other quality aspects. *Innovative Food Science and Emerging Technologies*, 6(3), 257–270. https://doi.org/10.1016/j.ifset.2005.04.001
- Nagashima, Y., Ebina, H., Tanaka, M., & Taguchi, T. (1993). Effect of high hydrostatic pressure on the thermal gelation of squid mantle meat. *Food Research International*, 26(2), 199–123. https://doi.org/ 10.1016/0963-9969(93)90067-S
- Ngadi, M. O., Latheef, M. B., & Kassama, L. (2012). Emerging technologies for microbial control in food processing. In J. I. Boye & Y. Arcand (Eds.), *Green technologies in food production* (pp. 363–411). Springer.
- Nierop Groot, M., Abee, T., & van Bokhorst-van de Veen, H. (2019). Inactivation of conidia from three *Penicillium* spp. isolated from fruit juices by conventional and alternative mild preservation technologies and disinfection treatments. *Food Microbiology*, *81*, 108– 114. https://doi.org/10.1016/j.fm.2018.06.004

- Nikzade, V., Sedaghat, N., Yazdi, F. T., Ghoddusi, H. B., & Saadatmand-Tarzjan, M. (2019). Development of shelf life kinetic model for fresh rainbow trout (*Oncorhynchus mykiss*) fillets stored under modified atmosphere packaging. *Journal of Food Science and Technology*, 56(2), 663–673. https://doi.org/10.1007/ s13197-018-3521-3
- Nilsson, L., Huss, H. H., & Gram, L. (1997). Inhibition of *Listeria monocytogenes* on cold-smoked salmon by nisin and carbon dioxide atmosphere. *International Journal of Food Microbiology*, *38*(2-3), 217–227. https://doi.org/10.1016/S0168-1605(97)00111-6
- Nishioka, M., Kanosue, F., Yabuta, Y., & Watanabe, f. (2011). Loss of vitamin B<sub>12</sub> in fish (round herring) meats during various cooking treatments. *Journal of Nutritional Science and Vitaminology*, 57(6), 432–436. https://doi.org/10.3177/jnsv.57.432
- Noseda, B., Islam, M. T., Eriksson, M., Heyndrickx, M., De Reu, K., Van Langenhove, H., & Devlieghere, F. (2012). Microbiological spoilage of vacuum and modified atmosphere packaged Vietnamese *Pangasius hypophthalmus* fillets. *Food Microbiology*, 30(2), 408–419. https://doi.org/10.1016/j.fm.2011.12.025
- Okozaki, T., Kakugawa, K., Yoneda, T., & Suzuki, K. (2000). Inactivation behavior of heat-resistant bacterial spores by thermal treatments combined with high hydrostatic pressure. *Food Science and Technology Research*, *6*(3), 204–207. https://doi.org/10.3136/fstr.6. 204
- Olatunde, O. O., & Benjakul, S. (2018). Nonthermal processes for shelf-life extension of seafoods: A revisit. *Comprehensive Reviews* in Food Science and Food Safety, 17(4), 892–904. https://doi.org/10. 1111/1541-4337.12354
- Olatunde, O. O., Benjakul, S., & Vongkamjan, K. (2020). Shelf-life of refrigerated Asian sea bass slices treated with cold plasma as affected by gas composition in packaging. *International Journal of Food Microbiology*, *324*, 108612. https://doi.org/10.1016/j. ijfoodmicro.2020.108612
- Olofsson, T. C., Ahrné, S., & Molin, G. (2007). The bacterial flora of vacuum-packed cold-smoked salmon stored at 7°C, identified by direct 16S rRNA gene analysis and pure culture technique. *Journal of Applied Microbiology*, *103*(1), 109–119. https://doi.org/10.1111/j.1365-2672.2006.03216.x
- Oms-Oliu, G., Martín-Belloso, O., & Sioliva-Fortuny, R. (2010). Pulsed light treatments for food preservation. A review. *Food and Bioprocess Technology*, *3*(1), 13–23. https://doi.org/10.1007/ s11947-008-0147-x
- Orsat, V., Raghavan, G. S. V., & Krishnaswamy, K. (2017). Microwave technology for food processing: An overview of current and future applications. In M. Regier, K. Knoerzer, & H. Schubert (Eds.), *The microwave processing of foods* (pp. 100–116). Woodhead Publishing.
- Ozer, N. P., & Demirci, A. (2006). Inactivation of Escherichia coli O157:H7 and Listeria monocytogenes inoculated on raw salmon fillets by pulsed UV-light treatment. International Journal of Food Science and Technology, 41(4), 354–360. https://doi.org/10.1111/j. 1365-2621.2005.01071.x
- Özoğul, Y. (Ed.). (2019). Innovative technologies in seafood processing (1st ed.). CRC Press. https://doi.org/10.1201/9780429327551
- Park, H., Hung, Y.-C., & Chung, D. (2004). Effects of chlorine and pH on efficacy of electrolyzed water for inactivating *Escherichia coli* O157:H7 and *Listeria monocytogenes*. *International Journal of Food Microbiology*, *91*(1), 13–18. https://doi.org/10.1016/S0168-1605(03) 00334-9



- Park, S. H., Balasubramaniam, V. M., Sastry, S. K., & Lee, J. (2013). Pressure-ohmic-thermal sterilization: A feasible approach for the inactivation of *Bacillus amyloliquefaciens* and *Geobacillus stearothermophilus* spores. *Innovative Food Science and Emerging Technologies*, 19, 115–123. https://doi.org/10.1016/j.ifset.2013.03. 005
- Park, S. Y., & Ha, S. D. (2015). Application of cold oxygen plasma for the reduction of *Cladosporium cladosporioides* and *Penicillium citrinum* on the surface of dried filefish (*Stephanolepis cirrhifer*) fillets. *International Journal of Food Science and Technology*, 50(4), 966–973. https://doi.org/10.1111/ijfs.12730
- Parlapani, F. F., Mallouchos, A., Haroutounian, S. A., & Boziaris, I. S. (2014). Microbiological spoilage and investigation of volatile profile during storage of sea bream fillets under various conditions. *International Journal of Food Microbiology*, 189, 153–163. https:// doi.org/10.1016/j.ijfoodmicro.2014.08.006
- Patil, S., Moiseev, T., Misra, N. N., Cullen, P. J., Mosnier, J. P., Keener, K. M., & Bourke, P. (2014). Influence of high voltage atmospheric cold plasma process parameters and role of relative humidity on inactivation of *Bacillus atrophaeus* spores inside a sealed package. *Journal of Hospital Infection*, 88(3), 162–169. https://doi.org/ 10.1016/j.jhin.2014.08.009
- Patterson, M. F. (2014). Food technologies: High pressure processing. In Y. Motarjemi (Ed.), *Encyclopedia of food safety* (Vol. 3, pp. 196– 201). Academic Press.
- Pedersen, S. J., Feyissa, A. H., Brøkner Kavli, S. T., & Frosch, S. (2016). An investigation on the application of ohmic heating of cold water shrimp and brine mixtures. *Journal of Food Engineering*, *179*, 28– 35. https://doi.org/10.1016/j.jfoodeng.2016.01.022
- Pedrós-Garrido, S., Condón-Abanto, S., Clemente, I., Beltrán, J. A., Lyng, J. G., Bolton, D., Brunton, N., & Whyte, P. (2018). Efficacy of ultraviolet light (UV-C) and pulsed light (PL) for the microbiological decontamination of raw salmon (*Salmo salar*) and food contact surface materials. *Innovative Food Science & Emerging Technologies*, 50, 124–131. https://doi.org/10.1016/j.ifset.2018.10.001
- Perni, S., Shama, G., & Kong, M. G. (2008). Cold atmospheric plasma disinfection of cut fruit surfaces contaminated with migrating microorganisms. *Journal of Food Protection*, 71(8), 1619–1625. https: //doi.org/10.4315/0362-028X-71.8.1619
- Pinto, L., Baruzzi, F., Cocolin, L., & Malfeito-Ferreira, M. (2020). Emerging technologies to control *Brettanomyces* spp. in wine: Recent advances and future trends. *Trends in Food Science & Technology*, 99, 88–100. https://doi.org/10.1016/j.tifs.2020.02.013
- Pollock, A. M., Pratap Singh, A., Ramaswamy, H. S., & Ngadi, M. O. (2017). Pulsed light destruction kinetics of *L. monocytogenes*. *LWT* - Food Science and Technology, 84, 114–121. https://doi.org/10.1016/ j.lwt.2017.05.040
- Pothuri, P., Marshall, D. L., & McMillin, K. W. (1996). Combined effects of packaging atmosphere and lactic acid on growth and survival of *Listeria monocytogenes* in crayfish tail meat at 4°C. *Journal of Food Protection*, 59(3), 253–256. https://doi.org/10.4315/ 0362-028X-59.3.253
- Prescott, S. C. (1927). The treatment of milk by electrical method. American Journal of Public health, 17(3), 221–223. https://doi.org/ 10.2105/ajph.17.3.221
- Provincial, L., Gil, M., Guillén, E., Alonso, V., Roncalés, P., & Beltrán, J. A. (2010). Effect of modified atmosphere packaging using different CO<sub>2</sub> and N<sub>2</sub> combinations on physical, chemical, microbiological and sensory changes of fresh sea bass (*Dicentrarchus labrax*)

fillets. International Journal of Food Science and Technology, 45(9), 1828–1836. https://doi.org/10.1111/j.1365-2621.2010.02341.x

- Provincial, L., Guillén, E., Gil, M., Alonso, V., Roncalés, P., & Beltrán, J. A. (2013). Survival of *Listeria monocytogenes* and *Salmonella Enteritidis* in sea bream (*Sparus aurata*) fillets packaged under enriched CO<sub>2</sub> modified atmospheres. *International Journal of Food Microbiology*, *162*(3), 213–219. https://doi.org/10. 1016/j.ijfoodmicro.2013.01.015
- Puligundla, P., Choi, S., & Mok, C. (2018). Microbial decontamination of *Gwamegi* (semi-dried Pacific saury) using corona discharge plasma jet, including physicochemical and sensory evaluation. *Journal of Aquatic Food Product Technology*, 27(3), 274–283. https://doi.org/10.1080/10498850.2017.1347592
- Qian, H., Li, W., Guo, L., Tan, L., Liu, H., Wang, J., Pan, Y., & Zhao, Y. (2020). Stress response of *Vibrio parahaemolyticus* and *Listeria monocytogenes* biofilms to different modified atmospheres. *Frontiers in Microbiology*, *11*, 23-23. https://doi.org/10.3389/fmicb.2020. 0002
- Rajasekaran, B., Subbiah, B., Stephen, N. M., Nagarajan, M., & Muniasamy, S. (2021). Design, fabrication, and validation of ohmic heater to process green mussel meat. *Journal of Food Processing* and Preservation, 45(6), e15511. https://doi.org/10.1111/jfpp.15511
- Rajkovic, A., Smigic, N., Uyttendaele, M., Medic, H., de Zutter, L., & Devlieghere, F. (2009). Resistance of *Listeria monocytogenes*, *Escherichia coli* O157:H7 and *Campylobacter jejuni* after exposure to repetitive cycles of mild bactericidal treatments. *Food Microbiology*, 26(8), 889–895. https://doi.org/10.1016/j.fm.2009.06.006
- Randell, K., Hattula, T., Skyttä, E., Sivertsvik, M., Bergslien, H., & Ahvenainen, R. (1999). Quality of filleted salmon in various retail packages. *Journal of Food Quality*, 22(5), 483–497. https://doi.org/ 10.1111/j.1745-4557.1999.tb00180.x
- Raso, J., & Barbosa-Cánovas, G. (2003). Nonthermal preservation of foods using combined processing techniques. *Critical Reviews* in Food Science and Nutrition, 43(3), 265–285. https://doi.org/10. 1080/10408690390826527
- Reddy, N. R., Solomon, H. M., Fingerhut, G., Rhodehamel, E. J., Balasubramaniam, V. M., & Palaniappan, S. (1999). Inactivation of *Clostridium botulinum* type E spores by high pressure processing. *Journal of Food Safety*, 19(4), 277–288. https://doi.org/10.1111/ j.1745-4565.1999.tb00252.x
- Regan, J. D., Trosko, J. E., & Carrier, W. L. (1968). Evidence for excision of ultraviolet-induced pyrimidine dimers from the DNA of human cells in vitro. *Biophysical Journal*, 8(3), 319–325. https://doi.org/10.1016/S0006-3495(68)86490-2
- Reineke, K., Mathys, A., & Knorr, D. (2011). The impact of high pressure and temperature on bacterial spores: Inactivation mechanisms of *Bacillus subtilis* above 500 MPa. *Journal of Food Science*, 76(3), M189–M197. https://doi.org/10.1111/j.1750-3841.2011.02066.x
- Ritz, M., Tholozan, J. L., Federighi, M., & Pilet, M. F. (2002). Physiological damages of *Listeria monocytogenes* treated by high hydrostatic pressure. *International Journal of Food Microbiology*, 79(1– 2), 47–53. https://doi.org/10.1016/S0168-1605(02)00178-2
- Roberts, P., & Hope, A. (2003). Virus inactivation by high intensity broad spectrum pulsed light. *Journal of Virological Methods*, *110*(1), 61–65. https://doi.org/10.1016/S0166-0934(03)00098-3
- Rode, T. M., Hovda, M. B., & Rotabakk, B. T. (2015). Favourable effects of soluble gas stabilisation and modified atmosphere for suppressing regrowth of high pressure treated Listeria innocua. *Food Control*, 51, 108–113. https://doi.org/10.1016/j.foodcont.2014.11.006

- Rodgers, S. (2016). Minimally processed functional foods: Technological and operational pathways. *Journal of Food Science*, *81*(10), R2309–R2319. https://doi.org/10.1111/1750-3841.13422
- Rodrigues, B. L., Alvares, T. S., Sampaio, G. S. L., Cabral, C. C., Araujo, J. V. A., Franco, R. M., Mano, S. B., & Conte, C. A., Jr. (2016). Influence of vacuum and modified atmosphere packaging in combination with UV-C radiation on the shelf life of rainbow trout (*Oncorhynchus mykiss*) fillets. *Food Control*, 60, 596–605. https://doi.org/10.1016/j.foodcont.2015.09.004
- Rong, C., Ling, Z., Huihui, S., & Qi, L. (2018). Characterization of microbial community in high-pressure treated oysters by highthroughput sequencing technology. *Innovative Food Science and Emerging Technologies*, 45, 241–248. https://doi.org/10.1016/j.ifset. 2017.11.001
- Rong, C., Ling, Z., & Qi, L. (2017). High pressure treatment changes spoilage characteristics and shelf life of pacific oysters (*Crassostrea* gigas) during refrigerated storage. Journal of Ocean University of China, 16(2), 351–355. https://doi.org/10.1007/s11802-017-3087-0
- Rosnes, J. T., & Skipnes, D. (2017). Minimal heat processing applied in fish processing. In D. Borda, A. I. Nicolau, & P. Raspor (Eds.), *Trends in fish processing technologies* (pp. 27–70). CRC Press.
- Rosnes, J. T., Skåra, T., & Skipnes, D. (2011). Recent advances in minimal heat processing of fish: Effects on microbiological activity and safety. *Food and Bioprocess Technology*, 4(6), 833–848. https: //doi.org/10.1007/s11947-011-0517-7
- Rotabakk, B. T., Birkeland, S., Jeksrud, W. K., & Sivertsvik, M. (2006). Effect of modified atmosphere packaging and soluble gas stabilization on the shelf life of skinless chicken breast fillets. *Journal of Food Science*, *71*(2), S124–S131. https://doi.org/10.1111/j.1365-2621. 2006.tb08915.x
- Rotabakk, B. T., Birkeland, S., Lekang, O. I., & Sivertsvik, M. (2008). Enhancement of modified atmosphere packaged farmed Atlantic Halibut (*Hippoglossus hippoglossus*) fillet quality by soluble gas stabilization. *Food Science and Technology International*, 14(2), 179–186. https://doi.org/10.1177/1082013208092051
- Rowan, N. J., MacGregor, S. J., Anderson, J. G., Fouracre, R. A., McIlvaney, L., & Farish, O. (1999). Pulsed-light inactivation of foodrelated microorganisms. *Applied and Environmental Microbiol*ogy, 65(3), 1312–1315. https://doi.org/10.1128/AEM.65.3.1312-1315. 1999
- Rutherford, T. J., Marshall, D., Andrews, L. S., Coggins, P. C., Wes Schilling, M., & Gerard, P. (2007). Combined effect of packaging atmosphere and storage temperature on growth of *Listeria monocytogenes* on ready-to-eat shrimp. *Food Microbiology*, 24(7-8), 703– 710. https://doi.org/10.1016/j.fm.2007.03.011
- Rørvik, L. M., Yndestad, M., & Skjerve, E. (1991). Growth of *Listeria* monocytogenes in vacuum-packed, smoked salmon, during storage at 4°C. International Journal of Food Microbiology, 14(2), 111–117. https://doi.org/10.1016/0168-1605(91)90097-9
- Saklani, P., Siddh, K. Das, S., Kumari Das, S., Ghosh, S. K., & Singh, S. M. (2019). Recent development of non-thermal cold plasma technology for safe and sustainable seafood processing: A review. *International Journal of Current Microbiology and Applied Sciences*, 8(11), 2459–2476. https://doi.org/10.20546/ijcmas.2019.811.284
- Salazar-González, C., San Martin-González, M. F., López-Malo, A., & Sosa-Morales, M. E. (2012). Recent studies related to microwave processing of fluid foods. *Food and Bioprocess Technology*, 5(1), 31– 46. https://doi.org/10.1007/s11947-011-0639-y

- Samal, S. (2017). Thermal plasma technology: The prospective future in material processing. *Journal of Cleaner Production*, 142, 3131– 3150. https://doi.org/10.1016/j.jclepro.2016.10.154
- Sampels, S. (2015). The effects of processing technologies and preparation on the final quality of fish products. *Trends in Food Science and Technology*, 44(2), 131–146. https://doi.org/10.1016/j.tifs.2015.04.003
- Sango, D. M., Abela, D., McElhatton, A., & Valdramidis, V. P. (2014). Assisted ultrasound applications for the production of safe foods. *Journal of Applied Microbiology*, *116*(5), 1067–1083. https://doi.org/ 10.1111/jam.12468
- Sastry, S. K. (2008). Ohmic heating and moderate electric field processing. Food Science and Technology International, 14(5), 419–422. https://doi.org/10.1177/1082013208098813
- Schellekens, M. (1996). New research issues in sous-vide cooking. Trends in Food Science and Technology, 7(8), 256–262. https://doi. org/10.1016/0924-2244(96)10027-3
- Shakila, R. J., Jeyasekaran, G., Vijayakumar, A., & Sukumar, D. (2009). Microbiological quality of sous-vide cook chill fish cakes during chilled storage (3°C). *International Journal of Food Science and Technology*, 44(11), 2120–2126. https://doi.org/10.1111/j. 1365-2621.2009.02047.x
- Sharma, G. (2010). Ultraviolet light. In Y. H. Hui (Ed.), Handbook of food science, technology and engineering (pp. 122-1–122-14). CRC Press.
- Sharma, R. R., & Demirci, A. (2003). Inactivation of Escherichia coli O157:H7 on inoculated alfalfa seeds with pulsed ultraviolet light and response surface modeling. Journal of Food Science, 68(4), 1448–1453. https://doi.org/10.1111/j.1365-2621.2003.tb09665.x
- Sheen, S., Huang, L., & Sommers, C. (2013). Survival of *Listeria mono-cytogenes*, *Escherichia coli* O157:H7, and *Salmonella* spp. on cat-fish fillets exposed to microwave heating in a continuous mode. *Journal of Food Science*, 77(8), E209-E214. https://doi.org/10.1111/j. 1750-3841.2012.02817.x
- Shiekh, K. A., & Benjakul, S. (2020). Effect of pulsed electric field treatments on melanosis and quality changes of Pacific white shrimp during refrigerated storage. *Journal of Food Processing and Preservation*, 44(1), e14292. https://doi.org/10.1111/jfpp.14292
- Shynkaryk, M. V., Pyatkovskyy, T., Yousef, A. E., & Sastry, S. K. (2020). In-situ monitoring of inactivation of *Listeria innocua* under high hydrostatic pressure using electrical conductivity measurement. *Journal of Food Engineering*, 285, 110087. https://doi.org/10.1016/ j.jfoodeng.2020.110087
- Silbande, A., Adenet, S., Smith-Ravin, J., Joffraud, J.-J., Rochefort, K., & Leroi, F. (2016). Quality assessment of ice-stored tropical yellowfin tuna (*Thunnus albacares*) and influence of vacuum and modified atmosphere packaging. *Food Microbiology*, 60, 62–72. https://doi.org/10.1016/j.fm.2016.06.016
- Sivertsvik, M. (2000). Use of soluble gas stabilisation to extend shelflife of salmon. Paper presented at the 29th WEFTA-Meeting, Greece.
- Sivertsvik, M. (2003). Active packaging in practice: Fish. In R. Ahvenainen (Ed.), *Novel food packaging techniques* (pp. 384–400). Woodhead Publishing in Food Science and Technology.
- Sivertsvik, M., & Birkeland, S. (2006). Effects of soluble gas stabilisation, modified atmosphere, gas to product volume ratio and storage on the microbiological and sensory characteristics of ready-toeat shrimp (*Pandalus borealis*). Food Science and Technology International, 12(5), 445–454. https://doi.org/10.1177/1082013206070171



- Sivertsvik, M., Jeksrud, W. K., & Rosnes, J. T. (2002). A review of modified atmosphere packaging of fish and fishery products - Significance of microbial growth, activities and safety. *International Journal of Food Science and Technology*, 37(2), 107–127. https://doi.org/ 10.1046/j.1365-2621.2002.00548.x
- Sivertsvik, M., Jeksrud, W. K., Vågane, Å., & Rosnes, J. T. (2004). Solubility and absorption rate of carbon dioxide into non-respiring foods: Part 1: Development and validation of experimental apparatus using a manometric method. *Journal of Food Engineering*, *61*(3), 449–458. https://doi.org/10.1016/S0260-8774(03)00167-5
- Sivertsvik, M., Rosnes, J. T., & Kleiberg, G. H. (2003). Effect of modified atmosphere packaging and superchilled storage on the microbial and sensory quality of Atlantic salmon (*Salmo salar*) fillets. *Journal of Food Science*, 68(4), 1467–1472. https://doi.org/10.1111/j. 1365-2621.2003.tb09668.x
- Smelt, J. P. P. M. (1998). Recent advances in the microbiology of high pressure processing. *Trends in Food Science and Technology*, 9(4), 152–158. https://doi.org/10.1016/S0924-2244(98)00030-2
- Sperber, W. H. (1983). Influence of water activity on foodborne bacteria - A review. *Journal of Food Protection*, 46(2), 142–150. https: //doi.org/10.4315/0362-028X-46.2.142
- Stammen, K., Gerdes, C., & Caporaso, F. (1990). Modified atmosphere packaging of seafood. *Critical Reviews in Food Science and Nutrition*, 29(5), 301–331. https://doi.org/10.1080/10408399009527530
- Stringer, S. C., Fernandes, M. A., & Metris, A. (2012). Safety of sous-vide foods: Feasibility of extending combase to describe the growth/survival/death response of bacterial foodborne pathogens between 40°C and 60°C. Final report on FSA project FS102028.
- Sun, B., Zhao, Y., Yu, J., Ling, J., Shang, H., & Liu, Z. (2017). The combined efficacy of superchilling and high CO<sub>2</sub> modified atmosphere packaging on shelf life and quality of swimming crab (*Portunus trituberculatus*). *Journal of Aquatic Food Product Technology*, 26(6), 655–664. https://doi.org/10.1080/10498850.2016.1252822
- Sørensen, J. S., Bøknæs, N., Mejlholm, O., & Dalgaard, P. (2020). Superchilling in combination with modified atmosphere packaging resulted in long shelf-life and limited microbial growth in Atlantic cod (*Gadus morhua* L.) from capture-based-aquaculture in Greenland. *Food Microbiology*, 88, 103405. https://doi.org/10. 1016/j.fm.2019.103405
- Tauscher, B. (1995). Pasteurization of food by hydrostatic high pressure: Chemical aspects. Zeitschrift für Lebensmittel- Untersuchung und -Forschung, 200(1), 3–13. https://doi.org/10.1007/BF01192901
- Teixeira, C., Fidalgo, L., Mendes, R., Costa, G., Cordeiro, C., Marques, A., Saraiva, J. A., & Nunes, M. L. (2014). Effect of high pressure processing in the quality of sea bass (*Dicentrarchus labrax*) fillets: Pressurization rate, pressure level and holding time. *Innovative Food Science and Emerging Technologies*, 22, 31–39. https: //doi.org/10.1016/j.ifset.2013.12.005
- Tholozan, J. L., Ritz, M., Jugiau, F., Federighi, M., & Tissier, J. P. (2000). Physiological effects of high hydrostatic pressure treatments on *Listeria monocytogenes* and *Salmonella typhimurium*. *Journal of Applied Microbiology*, 88(2), 202–212. https://doi.org/10. 1046/j.1365-2672.2000.00917.x
- Thostenson, E. T., & Chou, T.-W. (1999). Microwave processing: Fundamentals and applications. *Composites Part A: Applied Science and Manufacturing*, 30(9), 1055–1071. https://doi.org/10.1016/ S1359-835X(99)00020-2
- Tian, X., Yu, Q., Wu, W., & Dai, R. (2018). Inactivation of microorganisms in foods by ohmic heating: A review. *Journal of Food*

*Protection*, *81*(7), 1093–1107. https://doi.org/10.4315/0362-028X. JFP-17-343

- Timmermans, R. A. H., Nierop Groot, M. N., Nederhoff, A. L., van Boekel, M., Matser, A. M., & Mastwijk, H. C. (2014). Pulsed electric field processing of different fruit juices: Impact of pH and temperature on inactivation of spoilage and pathogenic microorganisms. *International Journal of Food Microbiology*, 173, 105– 111. https://doi.org/10.1016/j.ijfoodmicro.2013.12.022
- Tocmo, R., Krizman, K., Khoo, W. J., Phua, L. K., Kim, M., & Yuk, H.-G. (2014). *Listeria monocytogenes* in vacuum-packed smoked fish products: Occurrence, routes of contamination, and potential intervention measures. *Comprehensive Reviews in Food Science and Food Safety*, *13*(2), 172–189. https://doi.org/10.1111/1541-4337. 12052
- Truelstrup Hansen, L., Gill, T., Rontved, S. D., & Huss, H. H. (1996). Importance of autolysis and microbiological activity on quality of cold-smoked salmon. *Food Research International*, 29(2), 181–188. https://doi.org/10.1016/0963-9969(96)00003-8
- Truelstrup Hansen, L., & Huss, H. H. (1998). Comparison of the microflora isolated from spoiled cold-smoked salmon from three smokehouses. *Food Research International*, *31*(10), 703–711. https: //doi.org/10.1016/S0963-9969(99)00049-6
- Truong, B. Q., Buckow, R., Stathopoulos, C. E., & Nguyen, M. H. (2015). Advances in high-pressure processing of fish muscles. *Food Engineering Reviews*, 7(2), 109–129. https://doi.org/10.1007/ s12393-014-9084-9
- Tseng, S., Abramzon, N., Jackson, J. o., & Lin, W.-J. (2011). Gas discharge plasmas are effective in inactivating *Bacillus* and *Clostridium* spores. *Applied Microbiology and Biotechnology*, 93(6), 2563– 2570. https://doi.org/10.1007/s00253-011-3661-0
- Tsironi, T. N., Houhoula, D., & Taoukis, P. S. (2020). Hurdle technology for fish preservation. Aquaculture and Fisheries, 5(2), 64–71. https://doi.org/10.1016/j.aaf.2020.02.001
- Tsironi, T. N., Ntzimani, A., Gogou, E., Tsevdou, M., Semenoglou, I., Dermesonlouoglou, E., & Taoukis, P. S. (2019). Modeling the effect of active modified atmosphere packaging on the microbial stability and shelf life of gutted sea bass. *Applied Sciences*, 9(23), 5019. https: //doi.org/10.3390/app9235019
- Tsogas, A., Vatavali, K., Dimitriou, E., Badeka, A., Kontakos, S., & Kontominas, M. G. (2019). Combined effect of light salting and vacuum packaging on the microbiological, chemical, and sensory attributes of mullet fillets (*Mugil cephalus*) during refrigerated and frozen/refrigerated storage. *Journal of Food Processing and Preser*vation, 43(8), e14009. https://doi.org/10.1111/jfpp.14009
- Ulusoy, Ş., Alakavuk, D. Ü., Mol, S., & Coşansu, S. (2019). Effect of microwave cooking on foodborne pathogens in fish. *Journal of Food Processing and Preservation*, 43(8), e14045. https://doi.org/10. 1111/jfpp.14045
- Valø, T., Nordeng Jakobsen, A., & Lerfall, J. (2020). The use of atomized purified condensed smoke (PCS) in cold-smoke processing of Atlantic salmon - Effect on quality and microbiological stability of a lightly salted product. *Food Control*, *112*, 107155. https: //doi.org/10.1016/j.foodcont.2020.107155
- van Wyk, S., Farid, M. M., & Silva, F. V. M. (2018). SO2, high pressure processing and pulsed electric field treatments of red wine: Effect on sensory, Brettanomyces inactivation and other quality parameters during one year storage. *Innovative Food Science & Emerging Technologies*, 48, 204–211. https://doi.org/10.1016/j.ifset.2018. 06.016

- Varlet, V., Prost, C., & Serot, T. (2007). Volatile aldehydes in smoked fish: Analysis, methods, occurrence and mechanisms of formation. *Food Chemistry*, *105*(4), 1536–1556. https://doi.org/10.1016/j. foodchem.2007.03.041
- Varlet, V., Serot, T., Knockaert, C., Cornet, J., Cardinal, M., Monteau, F., Le Bizec, B., & Prost, C. (2007). Organoleptic characterization and PAH content of salmon (*Salmo salar*) fillets smoked according to four industrial smoking techniques. *Journal of the Science of Food and Agriculture*, 87(5), 847–854. https://doi.org/10.1002/jsfa. 2786
- Walkling-Ribeiro, M., Noci, F., Cronin, D. A., Riener, J., Lyng, J. G., & Morgan, D. J. (2008). Reduction of *Staphylococcus aureus* and quality changes in apple juice processed by ultraviolet irradiation, pre-heating and pulsed electric fields. *Journal of Food Engineering*, 89, 267–273. https://doi.org/10.1016/j.jfoodeng.2008.05.001
- Wang, T., Sveinsdóttir, K., Magnússon, H., & Martinsdóttir, E. (2008). Combined application of modified atmosphere packaging and superchilled storage to extend the shelf life of fresh cod (*Gadus morhua*) loins. *Journal of Food Science*, 73(1), S11–S19. https://doi. org/10.1111/j.1750-3841.2007.00590.x
- Wang, W., Li, M., & Li, Y. (2013). Modeling the thermoultrasound inactivation of *Vibrio parahaemolyticus* in raw peeled shrimps. *Journal of Food Protection*, 76(10), 1712–1718. https://doi.org/10. 4315/0362-028X.JFP-12-568
- Wang, J. J., Lin, T., Li, J. B., Liao, C., Pan, Y. J., & Zhao, Y. (2014). Effect of acidic electrolyzed water ice on quality of shrimp in dark condition. *Food Control*, 35(1), 207–212. https://doi.org/10.1016/j. foodcont.2013.07.005
- Wu, C. H., Yuan, C. H., Ye, X. Q., Hu, Y. Q., Chen, S. G., & Liu, D. H. (2014). A critical review on superchilling preservation technology in aquatic product. *Journal of Integrative Agriculture*, *13*(12), 2788– 2806. https://doi.org/10.1016/S2095-3119(14)60841-8
- Xuan, X.-t., Fan, Y.-f., Ling, J.-g., Hu, Y.-q., Liu, D.-h., Chen, S.-g., Ye, X.-q., & Ding, T. (2017). Preservation of squid by slightly acidic electrolyzed water ice. *Food Control*, 73(B), 1483–1489. https://doi. org/10.1016/j.foodcont.2016.11.013
- Yadav, B., Spinelli, A. C., Misra, N. N., Tsui, Y. Y., McMullen, L. M., & Roopesh, M. S. (2020). Effect of in-package atmospheric cold plasma discharge on microbial safety and quality of ready-to-eat ham in modified atmospheric packaging during storage. *Journal* of Food Science, 85(4), 1203–1212. https://doi.org/10.1111/1750-3841. 15072
- Yagiz, Y., Kristinsson, H. G., Balaban, M. O., & Marshall, M. R. (2007). Effect of high pressure treatment on the quality of rainbow trout (*Oncorhynchus mykiss*) and Mahi Mahi (*Coryphaena hippurus*). Journal of Food Science, 72(9), C509–C515. https://doi.org/10.1111/ j.1750-3841.2007.00560.x
- Yahia, E. M., Fadanelli, L., Mattè, P., & Brecht, J. K. (2019). Controlled atmosphere storage. In E. M. Yahia (Ed.), *Postharvest technology* of perishable horticultural commodities (pp. 439–479). Woodhead Publishing.

- Yildirim, S., Röcker, B., Pettersen, M. K., Nilsen-Nygaard, J., Ayhan, Z., Rutkaite, R., Radusin, T., Suminska, P., Marcos, B., & Coma, V. (2018). Active packaging applications for food. *Comprehensive Reviews in Food Science and Food Safety*, *17*(1), 165–199. https://doi. org/10.1111/1541-4337.12322
- Yogesh, K. (2016). Pulsed electric field processing of egg products: A review. *Journal of Food Science and Technology*, *53*(2), 934–945. https://doi.org/10.1007/s13197-015-2061-3
- Yu, P., Yan, C., Yang, F., Xu, Y., Jiang, Q., & Xia, W. (2018). Effect of high pressure processing on the quality and endogenous enzyme activities of grass carp (*Ctenopharyngodon idellus*) fillets stored at 4°C. *Journal of Aquatic Food Product Technology*, 27(10), 1093– 1105. https://doi.org/10.1080/10498850.2018.1534916
- Zander, K., & Feucht, Y. (2018). Consumers' willingness to pay for sustainable seafood made in Europe. *Journal of International Food* & Agribusiness Marketing, 30, 251–275. https://doi.org/10.1080/ 08974438.2017.1413611
- Zhang, B., Ma, L.-k., Deng, S.-g., Xie, C., & Qiu, X.-h. (2015). Shelflife of pacific white shrimp (*Litopenaeus vannamei*) as affected by weakly acidic electrolyzed water ice-glazing and modified atmosphere packaging. *Food Control*, *51*, 114–121. https://doi.org/10. 1016/j.foodcont.2014.11.016
- Zhao, L., Zhang, Z., Wang, M., Sun, J., Li, H., Malakar, P. K., Liu, H., Pan, Y., & Zhao, Y. (2018). New insights into the changes of the proteome and microbiome of shrimp (*Litopenaeus vannamei*) stored in acidic electrolyzed water ice. *Journal of Agricultural and Food Chemistry*, 66(19), 4966–4976. https://doi.org/10. 1021/acs.jafc.8b00498
- Zhou, Y., He, Q., & Zhou, D. (2017). Optimization extraction of protein from mussel by high-intensity pulsed electric fields. *Journal* of Food Processing and Preservation, 41(3), e12962. https://doi.org/ 10.1111/jfpp.12962
- Zeng, Q. Z., Thorarinsdottir, K. A., & Olafsdottir, G. (2005). Quality changes of shrimp (*Pandalus borealis*) stored under different cooling conditions. *Journal of Food Science*, 70(7), 459–466. https: //doi.org/10.1111/j.1365-2621.2005.tb11493.x
- Özpolat, E., & Patir, B. (2016). Determination of shelf life for sausages produced from some freshwater fish using two different smoking methods. *Journal of Food Safety*, *36*(1), 69–76. https://doi.org/10. 1111/jfs.12214

**How to cite this article:** Abel, N., Rotabakk, B. T., & Lerfall, J. Mild processing of seafood—A review. *Compr Rev Food Sci Food Saf*. 2021; 1–31 https://doi.org/10.1111/1541-4337.12876